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## (12) United States Patent

## Sheard et al.

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## (54) METHODS AND COMPOSITIONS FOR TARGETED PROTEIN DEGRADATION

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(2), (4) Date: Apr. 29, 2013

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PCT Pub. Date: **Dec. 15, 2011** 

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- (51) Int. Cl. C12N 15/00 (2006.01) C12N 15/81 (2006.01) C07K 14/415 (2006.01) C12N 15/62 (2006.01)

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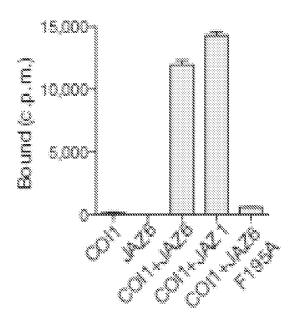
Primary Examiner — Anoop Singh (74) Attorney, Agent, or Firm — Perkins Coie LLP

## (57) ABSTRACT

Coronatine has been found to enhance binding of the JAZ1 degron to the *Arabidopsis* F-box protein COI1, and analysis of the JAZ1 degron sequence has resulted in the identification of specific peptide sequences that bind COI1 with high affinity in the presence of coronatine. Crystal structure analysis has determined that coronatine and JA-Ile enhance the interaction between COI1 and JAZ1 via binding to a specific binding pocket on COI1. Attachment of one or more JAZ1 peptide tags as disclosed herein to a target protein in a non-plant cell expressing *Arabidopsis* COI1 or a homolog thereof results in degradation of the target protein following addition of a molecule that binds the coronatine/JA-Ile binding pocket on COI1. Therefore, provided herein are compositions, methods, and kits for targeted protein degradation.

## 3 Claims, 25 Drawing Sheets

Figure 1



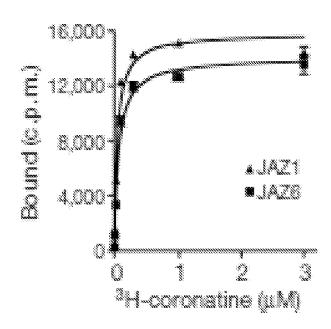
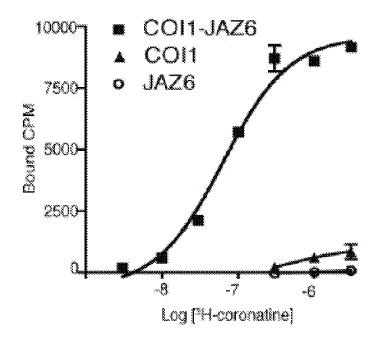


Figure 2



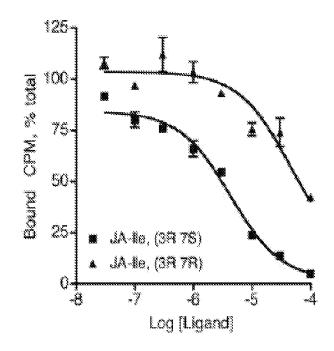


Figure 3

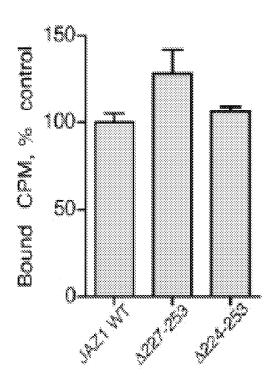


Figure 4

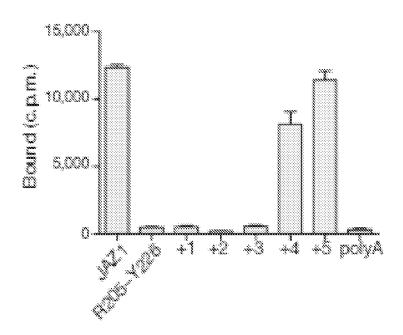


Figure 5

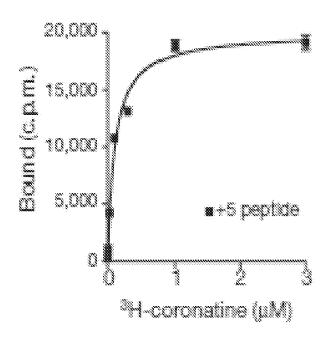


Figure 6

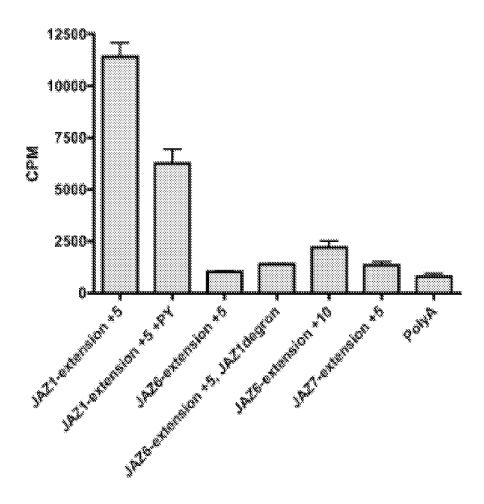
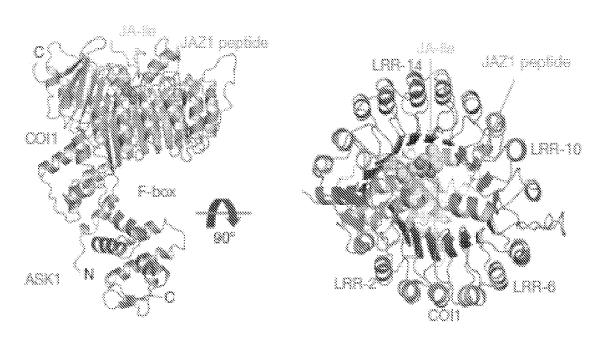


Figure 7

B. A.



C.

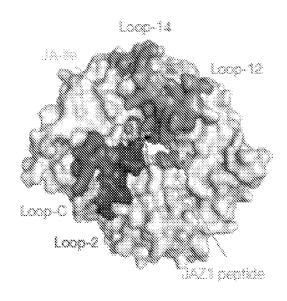
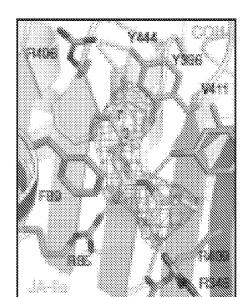
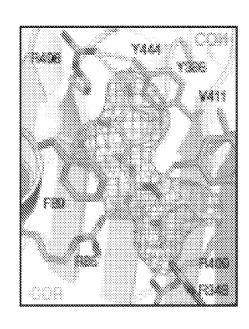


Figure 8



B.



C.

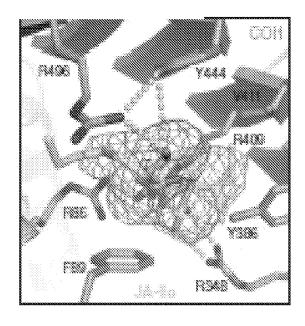


Figure 9

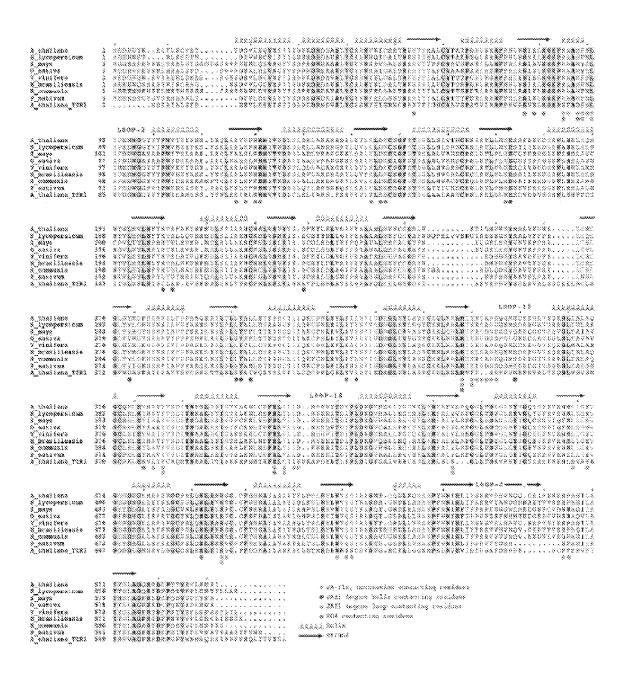
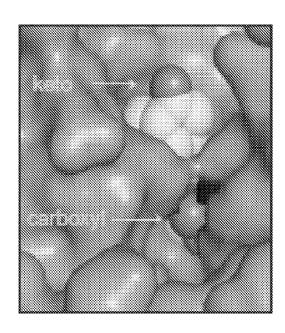


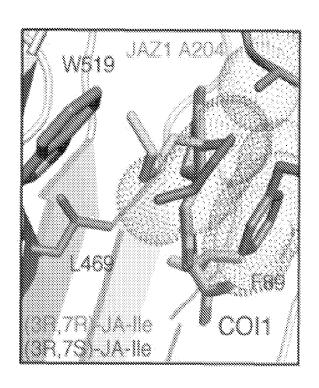
Figure 10



Mar. 29, 2016

Figure 11

A.



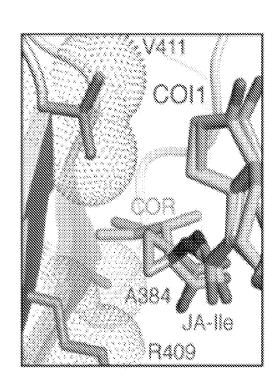
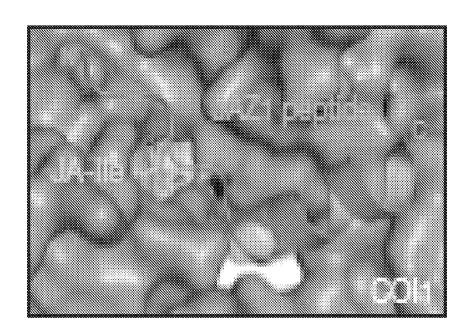


Figure 12



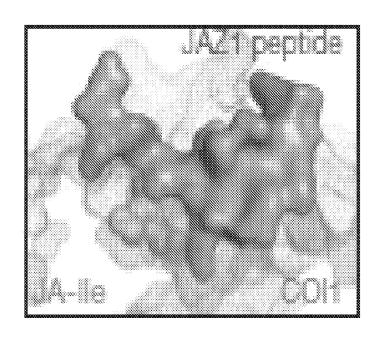
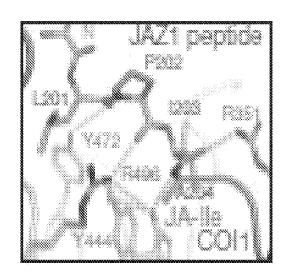
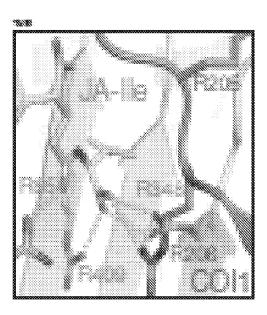


Figure 13



B.



C.

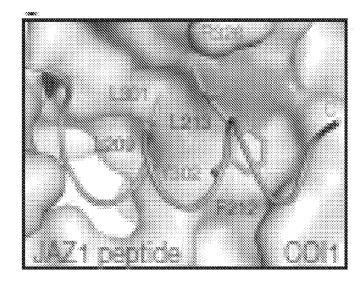
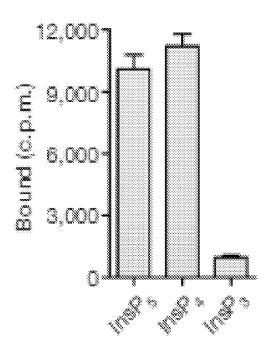


Figure 14



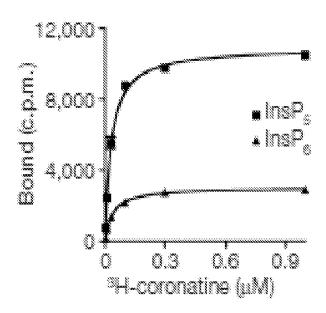


Figure 15

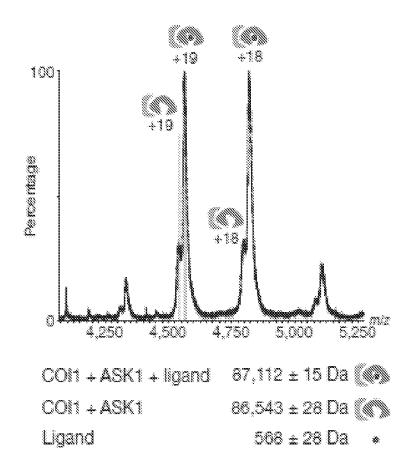


Figure 16

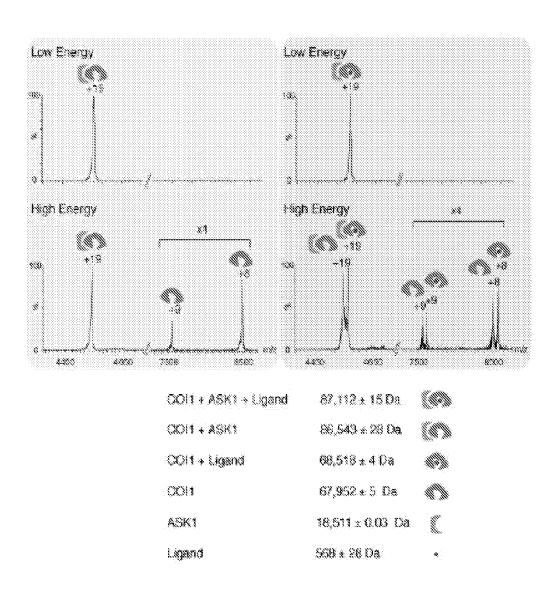


Figure 17

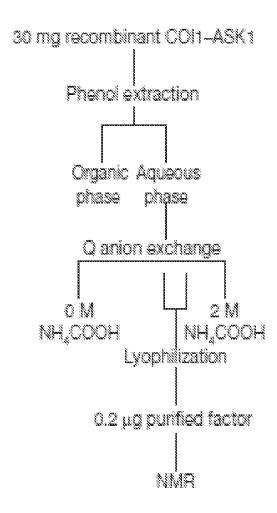


Figure 18

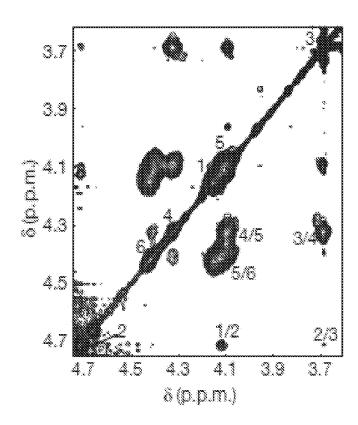


Figure 19

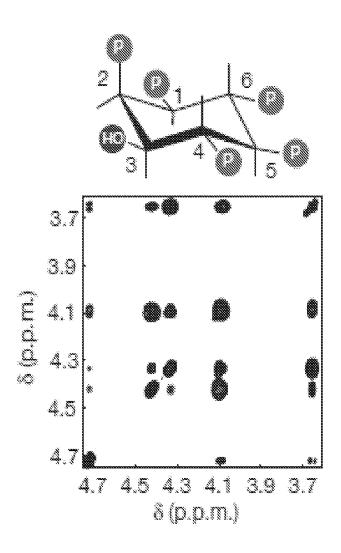
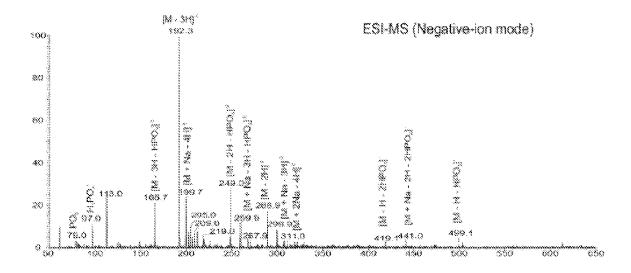


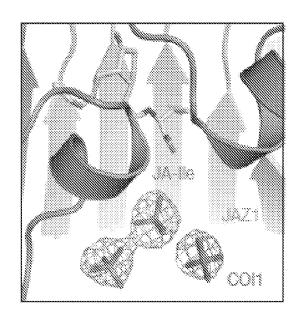
Figure 20



Mar. 29, 2016

Figure 21

A.



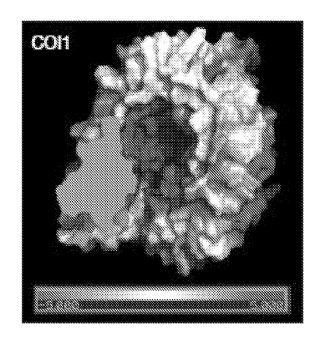


Figure 22

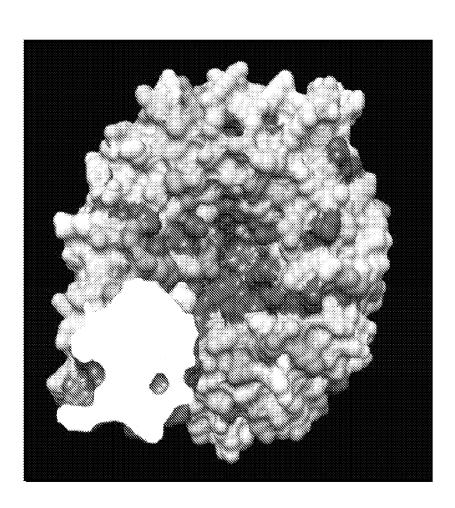
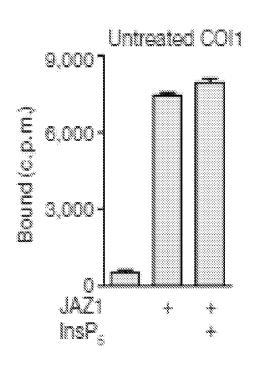
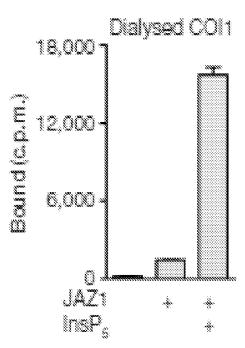


Figure 23







C.

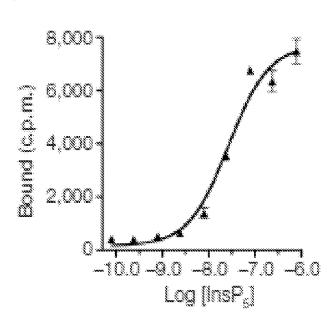


Figure 24

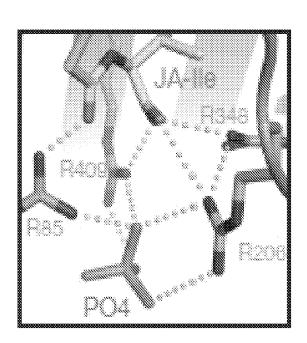
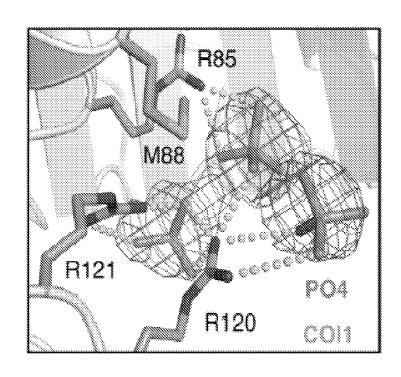
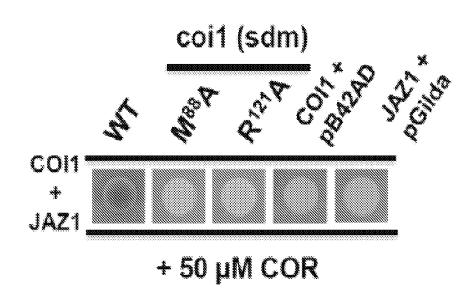


Figure 25





## METHODS AND COMPOSITIONS FOR TARGETED PROTEIN DEGRADATION

#### RELATED APPLICATIONS

The present application claims priority to U.S. Provisional Patent Application No. 61/352,758, filed Jun. 8, 2010, the disclosure of which is incorporated by reference herein in its entirety.

#### STATEMENT OF GOVERNMENT INTEREST

This invention was made with government support under grant number 2RO1CA107134 awarded by National Institutes of Health. The government has certain rights in the <sup>15</sup> invention.

## BACKGROUND

Reverse genetic approaches are a powerful laboratory tool 20 for determining the function of a target protein. The target protein is "knocked down," and cellular changes are observed in order to infer the normal function of the knocked down target. Methods for knocking down a target protein by manipulating DNA transcription and RNA translation are 25 well established. Among the most commonly used are gene knockout in whole animals and degradation of target mRNA using siRNA and shRNA techniques. However, delivery of RNA molecules can be cumbersome, and these methods often cannot achieve 100% efficiency.

Although several methods exist for knocking down a protein target at the transcription and translation levels, there are very few options for knocking down a protein target once the protein has been made. Such methods are desirable because most small molecule therapeutics operate by manipulating 35 proteins directly. Therefore, targeted degradation techniques that operate at the protein level provide the best tool for examining the potential effects of small molecule therapeutics.

The ideal protein knock down system would function at the 40 protein level, and would be capable of tightly controlling protein levels in a temporal manner. Temporal control of protein levels allows down-regulation of proteins that are essential for the full development of a system or pathway, and can be used to study dynamic biological functions which are 45 otherwise difficult to manipulate.

## **SUMMARY**

In certain embodiments, methods are provided for targeted 50 protein degradation. In certain of these embodiments, the protein targeted for degradation is tagged with one or more JAZ peptide tags as provided herein. The target protein is expressed in a non-plant host cell that also expresses the Arabidopsis protein COI1 or a homolog thereof, and a mol- 55 ecule that binds the COI1/JA-Ile binding pocket of COI1 is introduced into the cell to induce target protein degradation. In certain embodiments, the peptide tag comprises, consists of, or consists essentially of an amino acid sequence as set forth in SEQ ID NOs: 5, 6, 7, or 13. In certain embodiments, 60 the molecule that binds the COI1/JA-Ile binding pocket of COI1 is coronatine, JA, or a JA amino acid conjugate such as JA-Ile. In certain embodiments, an inositol pentakisphosphate cofactor may also be introduced into the cell. In certain embodiments, the non-plant host cell may be a eukaryotic cell such as a yeast or mammalian cell. In certain embodiments, the Arabidopsis protein COI1 or a homolog thereof may be

2

COI1 from Arabidopsis thaliana or Arabidopsis lyrata, or it may be a COI1 homolog from another plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or *Physcomitrella* patens. The target protein may be either an endogenous host cell protein or an exogenous protein. In those embodiments wherein the target protein is an endogenous host cell protein, the peptide tag may be attached to the target protein by introducing a DNA sequence encoding the peptide tag adjacent to the DNA sequence encoding the target protein in the host cell, such that the target protein is expressed with the peptide tag attached. In those embodiments wherein the target protein is an exogenous protein, the target protein may be introduced into the host cell via a DNA sequence encoding the target protein and the peptide tag. In certain embodiments, target protein degradation may be halted by deactivating the molecule that binds the COI1/JA-Ile binding pocket of COI1 or removing it from the cell. In other embodiments, target protein degradation may be halted by natural degradation of the molecule that binds the COI1/JA-Ile binding pocket of COI1.

In certain embodiments, methods are provided for targeted protein degradation. In certain of these embodiments, the protein targeted for degradation is tagged with one or more peptide tags comprising an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13. The target protein is expressed in a non-plant host cell that also expresses the Arabidopsis protein COI1 or a homolog thereof, and coronatine or JA-Ile is introduced into the cell to induce target protein degradation. In certain embodiments, an inositol pentakisphosphate cofactor may also be introduced into the cell. In certain embodiments, the non-plant host cell may be a eukaryotic cell such as a yeast or mammalian cell. In certain embodiments, the Arabidopsis protein COI1 or a homolog thereof may be COI1 from Arabidopsis thaliana or Arabidopsis lyrata, or it may be a COI1 homolog from another plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or Physcomitrella patens. The target protein may be either an endogenous host cell protein or an exogenous protein. In those embodiments wherein the target protein is an endogenous host cell protein, the peptide tag may be attached to the target protein by introducing a DNA sequence encoding the peptide tag adjacent to the DNA sequence encoding the target protein in the host cell, such that the target protein is expressed with the peptide tag attached. In those embodiments wherein the target protein is an exogenous protein, the target protein may be introduced into the host cell via a DNA sequence encoding the target protein and the peptide tag. In certain embodiments, target protein degradation may be halted by deactivating coronatine or JA-Ile or removing them from the cell. In other embodiments, target protein degradation may be halted by natural degradation of coronatine or JA-Ile.

In certain embodiments, methods are provided for targeted protein degradation in a non-plant host cell by fusing a target protein to a peptide tag as provided herein, introducing a DNA sequence encoding *Arabidopsis* COI1 or a homolog thereof into the host cell, culturing the host cell under conditions that result in the expression of the target protein and *Arabidopsis* COI1 or a homolog thereof, and introducing a molecule that binds the COI1/JA-Ile binding pocket of COI1 into the host cell. In certain embodiments, the peptide tag comprises, consists of, or consists essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13. In certain embodiments, the molecule that binds the COI1/JA-Ile binding pocket of COI1 is coronatine, JA, or a JA amino acid conjugate such as JA-Ile. In certain embodiments, an inositol pentakisphosphate cofactor may also be introduced

into the cell. In certain embodiments, the non-plant host cell may be a eukaryotic cell such as a yeast or mammalian cell. In certain embodiments, the Arabidopsis protein COI1 or a homolog thereof may be COI1 from Arabidopsis thaliana or Arabidopsis lyrata, or it may be a COI1 homolog from 5 another plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or Physcomitrella patens. The target protein may be either an endogenous host cell protein or an exogenous protein. In those embodiments wherein the target protein is an endogenous host cell protein, the peptide tag may be attached to the target protein by introducing a DNA sequence encoding the peptide tag adjacent to the DNA sequence encoding the target protein in the host cell, such that the target protein is expressed with the peptide tag attached. In those 15 embodiments wherein the target protein is an exogenous protein, the target protein may be introduced into the host cell via a DNA sequence encoding the target protein and the peptide tag. In certain embodiments, target protein degradation may be halted by deactivating the molecule that binds the COI1/ 20 JA-Ile binding pocket of COI1 or removing it from the cell. In other embodiments, target protein degradation may be halted by natural degradation of the molecule that binds the COI1/ JA-Ile binding pocket of COI1.

In certain embodiments, methods are provided for targeted 25 protein degradation in a non-plant host cell by fusing a target protein to a peptide tag comprising, consisting of, or consisting essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13, introducing a DNA sequence encoding Arabidopsis COI1 or a homolog thereof into the host cell, culturing the host cell under conditions that result in the expression of the target protein and Arabidopsis COI1 or a homolog thereof, and introducing coronatine or JA-Ile into the host cell. In certain embodiments, an inositol pentakisphosphate cofactor may also be introduced into the cell. In 35 certain embodiments, the non-plant host cell may be a eukaryotic cell such as a yeast or mammalian cell. In certain embodiments, the Arabidopsis protein COI1 or a homolog thereof may be COI1 from Arabidopsis thaliana or Arabidopsis lyrata, or it may be a COI1 homolog from another 40 plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or *Physcomitrella patens*. The target protein may be either an endogenous host cell protein or an exogenous protein. In those embodiments wherein the target protein is an 45 endogenous host cell protein, the peptide tag may be attached to the target protein by introducing a DNA sequence encoding the peptide tag adjacent to the DNA sequence encoding the target protein in the host cell, such that the target protein is expressed with the peptide tag attached. In those embodi- 50 ments wherein the target protein is an exogenous protein, the target protein may be introduced into the host cell via a DNA sequence encoding the target protein and the peptide tag. In certain embodiments, target protein degradation may be halted by deactivating coronatine or removing it from the cell. 55 In other embodiments, target protein degradation may be halted by natural degradation of coronatine.

In certain embodiments, methods are provided for targeted protein degradation in a host animal by introducing a DNA sequence encoding the target protein linked to a peptide tag as 60 provided herein and another DNA sequence encoding *Arabidopsis* COI1 or a homolog thereof, expressing the tagged target protein and COI1, and then administering a molecule that binds the COI1/JA-Ile binding pocket of COI1 to the animal. In certain embodiments, an inositol pentakisphosphate cofactor may also be introduced into the cell. In certain embodiments, the peptide tag comprises, consists of, or con-

4

sists essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13. In certain embodiments, the molecule that binds the COI1/JA-Ile binding pocket of COI1 is coronatine, JA, or a JA amino acid conjugate such as JA-Ile. In certain embodiments, the animal is a mammal, and in certain of these embodiments the animal is a mouse. In certain embodiments, the *Arabidopsis* protein COI1 or a homolog thereof may be COI1 from *Arabidopsis thaliana* or *Arabidopsis lyrata*, or it may be a COI1 homolog from another plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or *Physcomitrella patens*.

In certain embodiments, methods are provided for targeted protein degradation in a host animal by introducing a DNA sequence encoding the target protein linked to a peptide tag comprising, consisting of, or consisting essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13 and another DNA sequence encoding Arabidopsis COI1 or a homolog thereof, expressing the tagged target protein and COI1, and then administering coronatine or JA-Ile to the animal. In certain embodiments, an inositol pentakisphosphate cofactor may also be introduced into the cell. In certain embodiments, the animal is a mammal, and in certain of these embodiments the animal is a mouse. In certain embodiments, the Arabidopsis protein COI1 or a homolog thereof may be COI1 from Arabidopsis thaliana or Arabidopsis lyrata, or it may be a COI1 homolog from another plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or Physcomitrella

In certain embodiments, non-plant host cells are provided that comprise a DNA sequence encoding a target protein linked to a peptide tag comprising, consisting of, or consisting essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13 and another DNA sequence encoding Arabidopsis COI1 or a homolog thereof. In certain embodiments, the cells further comprise an inositol pentakisphosphate cofactor. In certain embodiments, the non-plant host cell is a eukaryotic cell, and in certain of these embodiments the non-plant host cell is a yeast or mammalian cell. In certain embodiments, the Arabidopsis protein COI1 or a homolog thereof may be COI1 from Arabidopsis thaliana or Arabidopsis lyrata, or it may be a COI1 homolog from another plant or moss such as rice, tomato, grape, poplar, castor oil, corn, rubber tree, pea, wild tobacco, soybean, sorghum, wheat, or Physcomitrella.

In certain embodiments, methods are provided for targeting an endogenous target protein in a non-plant host cell for coronatine- or JA-Ile-induced degradation by introducing a DNA sequence encoding a peptide tag comprising, consisting of, or consisting essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13 such that the DNA sequence is inserted adjacent to the gene encoding the endogenous target protein, and such that the target protein is expressed fused to the peptide tag.

In certain embodiments, peptides are provided for tagging a target protein for degradation, and in certain of these embodiments, the peptide tags comprise, consist of, or consist essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13. Also provided in certain embodiments are isolated nucleic acid sequences encoding these peptide tags, as well as the use of the peptide tags in tagging target proteins for degradation.

In certain embodiments, kits are provided for targeted protein degradation. In certain embodiments, these kits may include one or more of the following: an isolated nucleic acid encoding a peptide tag as provided herein, an isolated nucleic

acid encoding Arabidopsis COI1 or a homolog thereof, and a molecule that binds the COI1/JA-Ile binding pocket of COI1. In certain embodiments, the kit may further comprise a target protein or an isolated nucleic acid encoding a target protein, and/or an inositol pentakisphosphate cofactor. In certain 5 embodiments, the peptide tag comprises, consists of, or consists essentially of an amino acid sequence as set forth in SEQ ID NOs:5, 6, 7, or 13. In certain embodiments, the molecule that binds the COII/JA-Ile binding pocket of COI1 is coronatine, JA, or a JA amino acid conjugate such as JA-Ile. In 10 certain embodiments, the kit further comprises instructions for use and/or other printed materials.

#### BRIEF DESCRIPTION OF DRAWINGS

FIG. 1: A. Binding of <sup>3</sup>H-coronatine (300 nM) to COI1 alone, JAZ6 alone, JAZ6/COI1, JAZ1/COI1, and JAZ6F195A/COI1. B. Saturation binding of <sup>3</sup>H-coronatine to the complex of COI1/ASK1 in the presence of JAZ1 ( $\triangle$ , KD of  $68\pm15$  nM) and JAZ6 ( $\blacksquare$ , KD of  $48\pm13$  nM).

FIG. 2: A. Saturation binding of <sup>3</sup>H-coronatine (up to 3) μM) to the COI1/JAZ6 complex, isolated COI1, and isolated JAZ6. B. Competition binding of 100 nM <sup>3</sup>H—COR with (3R,7S)-JA-Ile and (3R,7R)-JA-Ile at a K, of  $1.8\pm0.6 \mu M$  and 18±19 μM, respectively.

FIG. 3: Binding of 300 nM <sup>3</sup>H—COR to JAZ1 proteins truncated immediately after (Δ227-253) or before (Δ224-253) the PY motif.

FIG. 4: Binding of 300 nM coronatine to COI1 in the presence of various JAZ1 degron derivative peptides with 30 systematic N-terminal extensions.

FIG. 5: Saturation binding of COI1/ASK1 and the JAZ1 degron +5 peptide. The peptide bound COI1 with a  $K_D$  of

various JAZ1 degron derivative peptides.

FIG. 7: A, B. Structure of Arabidopsis COI1 (green ribbon)/Ask1 receptor protein (grey ribbon) complex bound to JAZ1 degron peptide (orange ribbon) and (3R,7S)-JA-Ile in yellow space fill representation. C. Surface representation of 40 COI1 (grey) with loop 2 (blue), loop 12 (purple), and loop 14 (green) forming the JA-Ile binding pocket.

FIG. 8: A, B. Side view of JA-Ile and COR binding. Hormones are shown as stick models, along with positive F<sub>o</sub>-F<sub>c</sub> electron density, calculated before they were built into the 45 model (red mesh). Hydrogen bond and salt bridge networks are shown with yellow dashes. C. Top view of the JA-Ile pocket showing the F<sub>o</sub>-F<sub>c</sub> electron density, calculated before JA-Ile was built into the model (red mesh). The electron density of the pentenyl side chain of (3R,7S)-JA-Ile cannot 50 accommodate the (3R,7R)-JA-Ile side chain, which is constrained by the chiral configuration at the C7 position.

FIG. 9: Alignment of Arabidopsis thaliana TIR1 (SEQ ID NO:31) and various COI1 orthologs from select plant species (Arabidopsis thaliana, SEQ ID NO:15; Solanum lycopersi- 55 cum, SEQ ID NO:18; Zea mays, SEQ ID NO:23; Oryza sativa, SEQ ID NO:17; Vitis vinifera, SEQ ID NO:19; Hevea brasiliensis, SEQ ID NO:24; Ricinus communis, SEQ ID NO:22; Pisum sativum, SEQ ID NO:25). Secondary structure elements as determined in the crystal structure of the COI1/ 60 ASK1/JAZ1 degron peptide/JA-Ile complex are shown on top of the Arabidopsis thaliana COI1 sequence. Critical ligand-, phosphate-, and substrate-contacting residues are indicated by colored dots as described in the key.

FIG. 10: When bound to COI1, JA-Ile (yellow space fill) is 65 solvent accessible at both the keto group (top) and carboxyl group (bottom).

6

FIG. 11: A. Side view of the COI1 pocket accommodating the pentenyl side chain of (3R,7S)-JA-Ile (yellow stick). The pentenyl side chain of (3R,7R)-JA-Ile (magenta stick) is modeled on the structure of (3R,7S)-JA-Ile and rotated around the C7-C8 bond to minimize collision with JAZ1 Ala 204 and COI1 Phe 89. The electron clouds of nearby COI1 (green) and JAZ1 (orange) side chains, as well as the pentenyl side chain of (3R,7R)-JA-Ile (magenta) are shown in dot form. Ala 86 and Leu 91 of COI1 blocking the front view of the pocket are omitted for clarity. B. Side view of (3R,7S)-JA-Ile (yellow stick) and coronatine (cyan stick) showing a hydrophobic pocket that accommodates both the aliphatic isoleucine portion of JA-Ile and the cyclopropane ring of coronatine.

FIG. 12: A. Top view of the complete JAZ1 degron peptide (orange) bound to COI1 (green) and JA-Ile (yellow). B. Side view and surface representation of the JAZ peptide, which acts as a clamp to lock JA-Ile in the pocket.

FIG. 13:A. Interactions of the N-terminal region of the 20 JAZ1 degron with COI1 and JA-Ile. Hydrogen bonds are shown with yellow dashes. B. Structural role of the Arg 206 residue from the JAZ1 degron in coordinating the carboxyl group of JA-Ile with three basic residues of the COI1 ligand pocket floor. C. Top view of the amphipathic JAZ1 degron helix bound to COI1 with three hydrophobic residues of JAZ1 shown in stick representation (orange) and COI1 residues in colored surface representation.

FIG. 14: A. Binding assays performed with 100 nM <sup>3</sup>H-coronatine, dialyzed COI1, and 1 μM synthetic InsP<sub>5</sub>. B. Saturation binding of <sup>3</sup>H-coronatine to dialyzed COI1 in the presence of 1  $\mu$ M of InsP<sub>5</sub> and InsP<sub>6</sub> at a K<sub>d</sub> of 30±5 nM and 37±8 nM, respectively. All results are the mean±s.e. of up to three experiments performed in duplicate.

FIG. 15: Nano-electrospray mass spectrometry of the FIG. 6: Binding of coronatine to COI1 in the presence of 35 intact COI/ASK1 complex. Low-intensity charge series corresponds in mass to the cofactor-free COI1/ASK1 complex.

> FIG. 16: Structural mass spectrometry analysis of the COI1/ASK1 complex. A. Isolation at 4564 m/z of the 19+ charge state for tandem MS analysis (shown in blue in FIG. 24). B. MS/MS spectrum showing the dissociation products of ions isolated at 4588 m/z (shown in orange in FIG. 24).

FIG. 17: Optimized cofactor purification scheme.

FIG. 18: Proton TOCSY spectrum of the purified cofactor. Numbers along the diagonal indicate the positions of the six protons of Ins(1,2,4,5,6)P<sub>5</sub>. The cross-peaks corresponding to direct couplings are labeled. Other cross-peaks correspond to relaved connectivities.

FIG. 19: TOCSY spectrum of a synthetic Ins(1,2,4,5,6)P<sub>5</sub> as a standard.

FIG. 20: Mass spectrometry analysis of Ins(1,2,4,5,6)P<sub>5</sub> purified from recombinant COI1/ASK1.

FIG. 21: A. Islands of positive  $F_o$ – $F_c$  electron density (red mesh) below the hormone-binding pockets, which probably belong to inorganic phosphate molecules from the crystallization solutions that displace InsP<sub>5</sub> from the InsP<sub>5</sub>-binding site. B. Bottom view of a surface electrostatic potential representation of COI1 from positive (blue) to negative (red).

FIG. 22: Surface conservation mapping of COI1. Conservation mapping of COI1 surface based on sequences of COI1 orthologs from nine different species (A. thaliana, H. brasiliensis, R. communis, P. trichocarpa, V. cinifera, P. sativum, S. lycopersicum, Z. mays, O. sativa). Dark blue, light blue, and white surface regions indicate 98-100%, 60-98%, and <60% sequence conservation, respectively. The F-box portion of COI1 and its associated ASK1 are carved out for clarity reasons. Four phosphate molecules bound to COI1 are shown by red sticks. JAZ1 peptide and ASK1 are shown in grey.

FIG. 23: A. Binding of  $^3$ H-coronatine at 100 nM to a complex of COI1 and JAZ1, with the addition of 1  $\mu$ M synthetic Ins(1,2,4,5,6)P<sub>5</sub> (InsP<sub>5</sub>). B. With extensive dialysis to remove the co-purified InsP<sub>5</sub> cofactor, 100 nM  $^3$ H-coronatine no longer binds dialyzed COI1 in the presence of JAZ1. Synthetic InsP<sub>5</sub> rescues binding. C. InsP<sub>5</sub> rescues the binding of 100 nM  $^3$ H-coronatine to dialyzed COI1/ASK1 in the presence of JAZ1 with an EC<sub>50</sub> of 27±12 nM.

FIG. 24: Interwoven hydrogen bond network in the complex structure.

FIG. **25**: A. Close-up view of COI1 residues (green stick) in close vicinity to the inorganic phosphates occupying the InsP5 binding pocket (orange stick, with along with positive  $F_o$ – $F_c$  density in red mesh). Hydrogen bonds are shown with yellow dashes. B. Interaction of wild-type COI1 and COI1 mutants with JAZ1 detected by yeast two-hybrid assay.

## DETAILED DESCRIPTION

The following description of the invention is merely intended to illustrate various embodiments of the invention. As such, the specific modifications discussed are not to be construed as limitations on the scope of the invention. It will be apparent to one skilled in the art that various equivalents, 25 changes, and modifications may be made without departing from the scope of the invention, and it is understood that such equivalent embodiments are to be included herein.

The following abbreviations are used herein: COR, coronatine; cpm, counts per minute; JA, jasmonic acid; JA-Ile, 30 jasmonyl-L-isoleucine; JAZ, jasmonate ZIM-domain; LRR, leucine-rich repeat; ppm, parts per million; SCF, Skp1-Cullin-F box protein; sdm, site-directed mutants.

There are currently no satisfactory methods for knocking down a protein target in a temporally controlled manner using targeted protein degradation. Several previous attempts at developing such a method have utilized the ubiquitin proteasome system. However, all of these methods have serious drawbacks.

One method that employs the ubiquitin proteasome system 40 utilizes peptide-small molecule hybrids ("protacs"). These chimeric molecules encourage binding of a target protein to the  $\text{SCF}^{\beta \textit{TrCP}}$  complex, resulting in proteolysis of the target (Sakamoto 2000). However, the protacs used in these methods are not membrane-permeable, and therefore require 45 modifications to increase cell permeability or the use of microinjection. Another disadvantage of this system is offtarget effects that arise from "swamping out"  $\mathring{SCF}^{\beta \mathit{TrCP}}$  and blocking its ability to interact with endogenous targets. Thus, modification of endogenous SCF complexes has shown very 50 limited usefulness as a reverse genetics tool. Another previously developed method for targeted protein degradation utilized chimeric fusions consisting of the target protein and large binding domains of proteins targeted for degradation at SCF complexes. One such system paired the target protein 55 with the chi retinoblastoma (Rb)-binding domain derived from the human papillomavirus (HPV) oncoprotein E7 (Zhou 2000). This method showed some success in yeast and mammalian systems, but it is not inducible and requires the use of an obstructive large tag. A similar chimeric approach required 60 engineering of the F-box protein β-TrCP with a small, phosphorylated peptide that encourages binding of a particular target molecule to the F-box protein (Zhang 2003). However, this system required a great deal of engineering in order for the chimeric F-box protein to recognize only the target of interest and not endogenous targets. Therefore, it has not been widely used.

8

Recently, researchers in Japan have developed a method that utilizes the *Arabidopsis* and tomato auxin receptor TIR1 for rapid, hormone-induced protein degradation (Nishimura Nature Methods 2009). TIR1 integrates into the yeast and mammalian SCF scaffold and degrades proteins with the AUX/IAA tag, the natural substrate of TIR1. However, this tag is over 20 kDa, a large tag by biochemistry standards. A smaller tag has not yet been identified. As such, the TIR1 system has limited usefulness.

The phytohormone jasmonic acid (JA) and its metabolites regulate a wide spectrum of plant physiology, participating in normal development and growth processes as well as defense responses to environmental and pathogenic stressors. JA is activated upon specific conjugation to the amino acid L-isoleucine, which produces the highly bioactive hormonal signal (3R,7S)-jasmonyl-L-isoleucine (JA-Ile). Coronatine (COR) is a *Pseudomonas syringae* virulence factor that structurally mimics JA-Ile.

The discovery of coronatine-insensitive mutants enabled the identification of COI1 as a key player in the JA pathway. *Arabidopsis* COI1 is an F-box protein that functions as the substrate-recruiting module of the Skp1-Cullin-F box protein (SCF) ubiquitin E3 ligase complex. Like other E3 ligases, SCF<sup>COI</sup> is involved in the ubiquitination of proteins, which targets the proteins for subsequent degradation by the 26S proteasome.

Arabidopsis jasmonate ZIM-domain (JAZ) proteins such as JAZ1, JAZ6, JAZ7, and JAZ8 are SCF<sup>CO1</sup> substrate targets that associate with COI1 in a hormone-dependent manner. In the absence of hormone signal, the JAZ proteins actively repress the transcription factor MYC2, which binds to cisacting elements of jasmonate-response genes. In response to cues that upregulate JA-Ile synthesis, the hormone stimulates the specific binding of JAZ proteins to COI1, leading to poly-ubiquitylation and subsequent degradation of the JAZ proteins by the 26S proteasome. JAZ degradation relieves repression of MYC2 and probably other transcription factors, permitting the expression of JA-responsive genes. The role of COI1-mediated JAZ degradation in JA signaling is analogous to auxin signaling through the F-box protein TIR1, which promotes hormone-dependent turnover of the AUX/IAA transcriptional repressors.

The experimental results provided herein disclose the identification and characterization of a complete jasmonate receptor comprising COI1, a JAZ peptide, and inositol pentakisphosphate. Coronatine was found to bind the complexes of COI1/JAZ1 and COI1/JAZ6 complexes with high affinity while displaying minimal binding affinity for COI1, JAZ1, or JAZ6 alone. Crystal structure studies were used to identify a coronatine- and JA-Ile-binding pocket on COI1. Binding of these molecules to the COI1 binding pocket increases binding affinity between COI1 and JAZ1. Coronatine has been found to play a similar role in the binding of JAZ2, JAZ3, JAZ4, JAZ5, JAZ6, JAZ8, JAZ9, JAZ10, JAZ11, and JAZ12 to COI1. A single isoform of inositol pentakisphosphate (Ins(1, 2,4,5,6)P<sub>5</sub>) was found to co-purify with COI1, and functional assays showed that this molecule is a critical cofactor in the interaction between COI1 and JAZ proteins.

The COI1 binding region of JAZ proteins had previously been mapped to a carboxy-terminal Jas motif. To precisely map the minimal region of the Jas motif that it is required for high affinity binding of COI1 to JAZ1 in the presence of coronatine, the JAZ1 degron and various derivatives thereof were analyzed. This led to the identification of specific JAZ peptide tags with enhanced binding affinity for COI1 in the presence of coronatine.

Provided herein in certain embodiments are compositions comprising one or more JAZ peptide tags capable of binding *Arabidopsis* COI1 or a homolog thereof, as well as nucleic acids encoding these JAZ peptide tags, methods of using these peptide tags to mark a target protein for degradation, and the use of these peptide tags in various methods and kits for temporally controlled protein degradation.

In certain embodiments, the JAZ peptide tags provided herein consist of, consist essentially of, or comprise the JAZ1 degron as set forth in SEQ ID NO: 1. In other embodiments, the peptide tags consist of, consist essentially of, or comprise an amino acid sequence that corresponds to the JAZ1 degron of SEQ ID NO:1 but with one or more additions, deletions, or substitutions. In certain of these embodiments, the peptide tags consist of, consist essentially of, or comprise the amino acid sequence of SEQ ID NO: 1 but with one or more deletions from the C-terminal end. In certain other of these embodiments, the peptide tags consist of, consist essentially of, or comprise the amino acid sequence of SEQ ID NO:1 but 20 with one or more additions to the N-terminal end. In other embodiments, the peptide tags provided herein comprise a fragment of an Arabidopsis JAZ protein other than JAZ1, such as for example JAZ2, JAZ3, JAZ4, JAZ5, JAZ6, JAZ8, JAZ9, JAZ10, JAZ11, or JAZ12.

In certain preferred embodiments, the JAZ1 peptide tags disclosed herein consist of, consist essentially of, or comprise the amino acid sequence of SEQ ID NOs:5, 6, or 7. In other preferred embodiments, the peptide tags consist of, consist essentially of, or comprise an amino acid sequence that corresponds to the amino acid sequences of SEQ ID NOs:5, 6, or 7, but wherein one or more amino acid substitutions, additions, or deletions have been introduced into the sequence. For example, in certain embodiments the peptide tags may consist of, consist essentially of, or comprise the amino acid 35 sequence  $X_1X_2X_3X_4X_5RRX_8SLHRFLEKRKDRVX_{22}X_{23}$ X<sub>24</sub>X<sub>25</sub>X<sub>26</sub>X<sub>27</sub> (SEQ ID NO: 13), wherein X<sub>1</sub>, X<sub>2</sub>, X<sub>3</sub>, X<sub>4</sub>, and  $X_5$  are each independently absent or any amino acid,  $X_8$ is either Ala or Lys, and  $X_{22}, X_{23}, X_{24}, X_{25}, X_{26},$  and  $X_{27}$  are each independently absent or any amino acid. In certain of 40 these embodiments,  $X_1$  is either absent, Glu, or Val;  $X_2$  is either absent, Leu, or Glu; X<sub>3</sub> is either absent, Pro, or Arg; X<sub>4</sub> is either absent or Ile; and  $X_5$  is either absent or Ala. In certain embodiments,  $X_{22}$  is either absent or Thr;  $X_{23}$  is either absent or Ser;  $X_{24}$  is either absent or Lys;  $X_{25}$  is either absent or Ala; 45  $X_{26}$  is either absent or Pro; and  $X_{27}$  is either absent or Tyr.

The small, unobtrusive peptide tags provided herein are superior to the large chimeric tags used in previously developed methods for targeted protein destruction because they are less likely to interfere with protein function, complex 50 formation, and subcellular localization. However, in certain embodiments, the peptide tags disclosed herein may comprise a longer sequence, such as for example the full-length JAZ1 polypeptide sequence as set forth in SEQ ID NO: 14 or the full-length JAZ10 polypeptide sequence set forth in SEQ 55 ID NO:29. In other of these embodiments, the peptide tag may comprise the full-length JAZ2, JAZ3, JAZ4, JAZ5, JAZ6, JAZ8, JAZ9, JAZ11, or JAZ12 polypeptide sequence.

COI1 binding to a protein comprising a JAZ peptide tag as results in degradation of the protein. As disclosed herein, 60 binding of coronatine or JA-Ile to a specific binding pocket in COI1 enhances the interaction of COI1 and the JAZ peptide. Thus, targeted protein degradation can be accomplished by tagging a target protein with a JAZ peptide tag as provided herein, then contacting the protein with COI1 in the presence 65 of a molecule that binds to the coronatine/JA-Ile binding pocket of COI1.

10

As such, provided herein in certain embodiments are methods for targeted protein degradation that utilize one or more of 1) *Arabidopsis* COI1 or a homolog thereof, 2) a molecule that binds the COI1/JA-Ile binding pocket in COI1, 3) one or more JAZ peptide tags, and, optionally, 4) an inositol pentakisphosphate cofactor. Also provided herein are compositions and kits for carrying out these methods.

The crystal structure analysis provided herein shows that coronatine and JA-Ile interact with a specific set of residues in the COI1 binding pocket that includes R85, A86, F89, L91, R348, E350, A384, Y386, R409, V411, Y444, LA69, R496, and W519 of SEQ ID NO:15. Therefore, a "molecule that binds the COI1/JA-Ile binding pocket of COI1" as used herein refers to a molecule that interacts with one or more of these residues, and more preferably with all fourteen of these residues. As used herein, a molecule "interacts" with a particular COI1 binding pocket residue if, when the molecule is bound to COI1, any portion of the molecule resides within a molecular distance that is within the hydrogen bond or Van der Waals interaction radius (approximately 2.5 to 4 Å) of the residue. In certain embodiments of the compositions, methods, and kits provided herein, a molecule that binds the COI1/ JA-Ile binding pocket of COI1 is coronatine. In other embodiments, the molecule is JA or a JA-amino acid conjugate such as JA-Ile, JA-L-leucine, JA-L-valine, or JA-L-alanine.

In those embodiments of the methods, compositions, and kits provided herein that utilize or comprise an inositol pentakisphosphate cofactor, the inositol pentakisphosphate cofactor may be inositol-1,2,4,5,6-pentakisphosphate. In other embodiments, the cofactor may be another molecule of the myo-inositol family, such as inositol-1,4,5,6-tetrakisphosphate.

In certain embodiments of the methods provided herein, one or more JAZ peptide tags are attached to or incorporated into a target protein. In the presence of molecule that binds the COI1/JA-Ile binding pocket of COI1, the JAZ peptide tag (and hence the target protein) binds to *Arabidopsis* COI1 or a homolog thereof with high affinity, resulting in target protein degradation.

Unlike previously developed methods for targeted protein destruction, the methods provided herein utilize small molecules that are membrane-permeable and require minimal engineering. The methods provided herein are rapidly inducible and easily reversible, and they do not require the use of large, obstructive tags. Thus, these methods represent a cheap, simple means for targeted protein destruction in vivo that is significantly superior to previously developed methods.

In certain embodiments of the methods disclosed herein, the target protein is tagged with one or more JAZ peptide tags as disclosed herein. JAZ peptide tags may be attached to a target protein using any methods known in the art. For example, in certain embodiments the peptide tag may be attached to the target protein prior to target protein expression. In these embodiments, a DNA sequence encoding the peptide tag is introduced into the host cell adjacent to the DNA sequence encoding the target protein, such that the peptide tag is expressed as part of the target protein. Methods for introducing a DNA sequence encoding a peptide tag into a cell and expressing a protein attached to the peptide tag are well known in the art. In other embodiments, the peptide tag may be attached to the target protein after the target protein has been expressed.

In certain embodiments of the methods disclosed herein, the target protein is tagged with a single peptide tag. In other embodiments, the target protein is tagged with two or more peptide tags. In those embodiments wherein the target protein

is tagged with two or more peptide tags, the peptide tags may have the same or different sequences.

Previously developed targeted protein destruction systems have generally utilized chimeric mammalian F-box proteins. However, the methods disclosed herein utilize the *Arabidop*sis F-box protein COI1 or plant or moss homologs thereof. In certain embodiments, the methods disclosed herein utilize Arabidopsis thaliana or Arabidopsis lyrata COI1 comprising the amino acid sequence set forth in SEQ ID NOs: 15 and 16, respectively. Specific plant or moss homologs that may be 10 utilized include those from rice (Oryza sativa, SEQ ID NO:17), tomato (Solanum lycopersicum, SEQ ID NO:18), grape (Vitis vinifera, SEQ ID NO:19), poplar (Populus trichocarpa, SEQ ID NO:20 and SEQ ID NO:21), castor oil (Ricinis communis, SEQ ID NO:22), corn (Zea mays, SEQ ID NO:23), rubber tree (Hevea brasiliensis, SEQ ID NO:24), pea (Pisum sativum, SEQ ID NO:25), wild tobacco (Nicotiana attenuata, SEQ ID NO:26), soybean (Glycine max, SEQ ID NO:27), sorghum (Sorghum bicolor, SEQ ID NO:28), wheat, able to mammalian proteins because their high specificity for plant substrates minimizes competition and unwanted degradation of endogenous targets.

In certain embodiments, an Arabidopsis COI1 polypeptide or a homolog thereof may be introduced directly into a cell. In 25 other embodiments, one or more genes encoding Arabidopsis COI1 or a homolog thereof may be introduced into a cell such that the cell expresses Arabidopsis COI1 or a homolog thereof. In these embodiments, the one or more genes encoding Arabidopsis COI1 or a homolog thereof may be intro- 30 duced into a cell via any method known in the art, including transformation or transient or stable transfection using viral and non-viral vectors. In certain embodiments, the one or more genes may be introduced using a viral vector such as an adenoviral, retroviral, lentiviral, or baculoviral vector. Pro- 35 vided herein in certain embodiments are viral and non-viral vectors comprising a DNA sequence encoding Arabidopsis COI1 or a homolog thereof, as well as methods of using these vectors to achieve expression of Arabidopsis COI1 or a homolog thereof in a non-plant host cell. In certain embodi- 40 ments, a DNA sequence encoding Arabidopsis COI1 or a homolog thereof may be incorporated into the host cell genome. In other embodiments, Arabidopsis COI1 or a homolog thereof may be expressed from a vector that is not incorporated into the host cell genome. The DNA sequence 45 encoding Arabidopsis COI1 or a homolog thereof can be placed under the control of an endogenous promoter that is naturally present in a host cell, or it may be placed under the control of an exogenous promoter that has been introduced into the cell in conjunction with the COI1 or homolog 50 sequence. In certain embodiments, Arabidopsis COI1 or a homolog thereof may be constitutively expressed in the host cell. In other embodiments, Arabidopsis COI1 or a homolog thereof may be expressed in a regulated manner. In certain of these embodiments, the DNA sequence encoding COI1 or a 55 homolog thereof may be placed under the control of an inducible promoter, such as a chemically-regulated promoter or physically-regulated promoter. In these embodiments, the inducible promoter may provide an additional layer of control over activation of targeted protein degradation.

In certain embodiments of the methods provided herein, *Arabidopsis* COI1 or a homolog thereof functions in conjunction with endogenous proteins to form a functional SCF<sup>CO1</sup> E3 ligase in the non-plant cell into which COI1 has been introduced. For example, exogenous COI1 that has been 65 introduced into a cell may function in combination with endogenous SKP1 to form a functional E3 ligase. Since SKP1

12

is highly conserved among species, the resultant complex is expected to be functional in most non-plant cell types. Nonetheless, in certain embodiments, one or more DNA sequences encoding SKP1 or other E3 ligase or ubiquitin pathway components may be introduced into a cell along with the one or more genes encoding *Arabidopsis* COI1 or homologs thereof. These other DNA sequences may be incorporated as part of the same vector as *Arabidopsis* COI1 or a homolog thereof, or they may be introduced via one or more separate vectors.

In certain embodiments, one or more modifications may be incorporated into *Arabidopsis* COI1 or a homolog thereof to enhance the interaction between COI1 or a homolog thereof and endogenous E3 ligase or other ubiquitin pathway proteins. These modifications may include one or more additions, substitutions, or deletions to the encoded COI1 or homolog sequence. Modifications may also include the addition of one or more peptide tags or the introduction of one or more covalent or non-covalent modifications.

In the methods disclosed herein, a molecule that binds the or Physcomitrella patens. COI1 and its homologs are prefer- 20 COI1/JA-Ile binding pocket of COI1 modulates the interaction between Arabidopsis COI1 or a homolog thereof and a target protein tagged with a JAZ peptide tag. In certain embodiments, the peptide tag will only bind Arabidopsis COI1 or a homolog thereof in the presence of the molecule that binds the COI1/JA-Ile binding pocket of COI1. In other embodiments, the peptide tag may bind Arabidopsis COI1 or a homolog thereof with very low affinity in the absence of the molecule, but do so with a significantly higher affinity in the presence of the molecule. The addition to or removal of the molecule that binds the COI1/JA-Ile binding pocket of COI1 from a cell provides a precise mechanism whereby targeted protein degradation can be turned on and off. For example, coronatine can be introduced into a cell to induce specific degradation, then withdrawn to halt degradation. Thus, the methods provided herein allow for precise temporal control of target protein degradation.

A molecule that binds the COI1/JA-Ile binding pocket of COI1 may be introduced into a cell and/or animal via any administration pathway known in the art. For example, the molecule can be administrated to a whole animal model via oral or parenteral administration routes. Introduction of the molecule into a host cell may be carried out via a single administration or via multiple administrations over a set time period. In certain embodiments, the molecule may be steadily administered to a host cell or animal over a set period of time. Withdrawal of the molecule from the host cell may occur via natural degradation of the molecule or by active removal or deactivation, such as by introduction of a neutralizing molecule that degrades or inactivates the molecule.

The methods and compositions disclosed herein may be utilized for targeted protein degradation in any non-plant host cell, including for example eukaryotic cells such as yeast or mammalian cells. Accordingly, provided herein in certain embodiments are non-plant host cells comprising DNA sequences encoding a target protein, a JAZ peptide tag, and *Arabidopsis* COI1 or a homolog thereof. Also provided herein are cell culture systems comprising such non-plant host cells.

The methods and compositions disclosed herein may also be utilized for targeted protein degradation in whole animals and animal models. Therefore, in certain embodiments these animals and animal models are also provided herein. In certain embodiments, the animal model is a mammalian animal model, and in certain of these embodiments the mammal is a rat or mouse, such as for example a knockout mouse model.

A target protein to be tagged for degradation using the compositions and methods disclosed herein may be an endog-

enous host cell protein. Alternatively, the target protein may be an exogenous protein that has been stably or transiently introduced into the host cell. In those embodiments of the methods disclosed herein where the methods are carried out in an animal model and wherein the target protein is an 5 exogenous protein, DNA sequences encoding the target protein, peptide tag, and Arabidopsis COI1 or a homolog thereof may be introduced into the animal using standard protein knockout methods known in the art. For example, the DNA sequences may be introduced into an embryonic stem cell under the control of one or more exogenous or endogenous promoters. This stem cell may be introduced into animal blastocysts, followed by selection for progeny that are homozygous for the introduction. Alternatively, the DNA sequences may be introduced into the animal at a later stage 15 via one or more non-plant cells comprising each of these DNA sequences or by direct transfection of one or more animal cells. In those embodiments wherein the methods are carried out in an animal model and the target protein is an endogenous protein, DNA sequences encoding the peptide 20 tag and Arabidopsis COI1 or a homolog thereof may be introduced into the animal by transfection of one or more animal cells. In these embodiments, the DNA sequence encoding the peptide tag is introduced in such a manner that is it is expressed as a fusion tag to the target protein.

In certain embodiments of the methods provided herein, targeted protein degradation is accomplished by the steps of 1) tagging a target protein with one or more peptide tags comprising, consisting of, or consisting essentially of an amino acid sequence selected from the group consisting of 30 SEQ ID NOs:5, 6, 7, and 13, b) expressing the target protein in a non-plant host cell, c) expressing *Arabidopsis* protein COI1 or a homolog thereof in the host cell, and 4) introducing coronatine or a jasmonic acid-amino acid conjugate into the host cell, resulting in degradation of the target protein.

In certain embodiments of the methods provided herein, targeted protein degradation in a non-plant host cell is accomplished by the steps of 1) attaching a peptide tag comprising, consisting of, or consisting essentially of an amino acid sequence selected from the group consisting of SEQ ID NOs: 40 5, 6, 7, and 13 to a target protein, 2) introducing a DNA sequence encoding *Arabidopsis* COI1 or a homolog thereof into the host cell, 3) culturing the host cell under conditions that result in the expression of *Arabidopsis* COI1 or a homolog thereof, and 4) introducing coronatine or a jasmonic 45 acid-amino acid conjugate into the host cell, resulting in degradation of the target protein.

In certain embodiments, kits are provided for carrying out targeted protein degradation in a non-plant host cell. In certain embodiments, these kits comprise one or more of the 50 following components: an isolated nucleic acid encoding a JAZ peptide tag as disclosed herein, an isolated nucleic acid encoding *Arabidopsis* COI1 or a homolog thereof, a molecule that binds to the coronatine/JA-Ile binding pocket of COI1, and/or an inositol pentakisphosphate cofactor. In certain 55 embodiments, the kit may further comprise a target protein or an isolated nucleic acid encoding a target protein. In certain embodiments, these kits further comprise instructions for usage.

The following examples are provided to better illustrate the claimed invention and are not to be interpreted as limiting the scope of the invention. To the extent that specific materials are mentioned, it is merely for purposes of illustration and is not intended to limit the invention. One skilled in the art may develop equivalent means or reactants without the exercise of 65 inventive capacity and without departing from the scope of the invention. It will be understood that many variations can

14

be made in the procedures herein described while still remaining within the bounds of the present invention. It is the intention of the inventors that such variations are included within the scope of the invention.

## **EXAMPLES**

## Example 1

Effect of Coronatine on COI1 Binding to JAZ1 and JAZ6

Various radioligand binding experiments were performed to quantify the interaction between tritium (<sup>3</sup>H)-labeled coronatine and COI1/JAZ1 or COI1/JAZ6.

(<sup>3</sup>H)-labeled coronatine was synthesized by Amersham. Full-length *Arabidopsis thaliana* COI1 and ASK1 were coexpressed as a glutathione S-transferase (GST) fusion protein and an untagged protein, respectively, in Hi5 suspension insect cells. The COI1/ASK1 complex was isolated from the soluble cell lysate by glutathione affinity chromatography. After on-column tag cleavage by tobacco etch virus protease, the complex was further purified by anion exchange and gel filtration chromatography and concentrated by ultrafiltration to 12-18 mg ml<sup>-1</sup>. Full-length JAZ substrate proteins were expressed as 6×His-fusion proteins in *Escherichia coli* and purified on Ni-NTA resin with subsequent dialysis into 20 mM Tris-HCl, pH 8.0, 200 mM NaCl, and 10% glycerol.

Radioligand binding was assayed on purified proteins, with 2 mg COII/ASK1 complex and JAZ proteins at a 1:3 molar ratio. Reactions were prepared in 100 ml final volume and in a binding buffer containing 20 mM Tris-HCl 200 mM NaCl, and 10% glycerol. Saturation binding experiments 35 were conducted with serial dilutions of 3H-coronatine in binding buffer. Nonspecific binding was determined in the presence of 300 mM coronatine. Competition binding experiments were conducted with serial dilutions of JA-Ile in the presence of 100 nM <sup>3</sup>H coronatine with nonspecific binding determined in the presence of 300 mM coronatine. Total binding was determined in the presence of vehicle only. Twopoint binding experiments were performed in the presence of 100 nM or 300 nM <sup>3</sup>H-coronatine with nonspecific binding determined in the presence of 300 mM coronatine. Following incubation with mixing at 4° C., all samples were collected with a cell harvester (Brandel, Gaithersburg, Md.) on polyethyleneimine (Sigma)-treated filters. Samples were incubated in liquid scintillation fluid for >1 hour before counting with a Packard Tri-Carb 2200 CA liquid scintillation analyzer (Packard Instrument Co.). Saturation binding experiments were analyzed by nonlinear regression, competition binding experiments by nonlinear regression with  $K_i$  calculation as described previously (Cheng 1973), and concentration-response data by sigmoidal dose-response curve fitting, all using GraphPad Prism version 4.00 for MacOSX.

<sup>3</sup>H-coronatine showed no appreciable binding affinity for COI1, full-length JAZ1, or full-length JAZ6 alone (FIG. 1A), but bound to the complex of COI1/JAZ1 with a  $K_D$  of 48 nM and to the complex of COI1/JAZ6 with a  $K_D$  of 68 nM (FIG. 1B). Binding of coronatine to the COI1/JAZ6 complex reached the level of saturation at 300 nM. Binding to COI1 alone at the same concentration elicited <2% specific binding (FIG. 2A). The highly active (3R,7S) isomer of JA-Ile was found to compete with coronatine for binding to the COI1-JAZ6 complex with an inhibition constant ( $K_i$ ) of 1.8 μM, while the less active (3R,7R) isomer competed with a  $K_i$  of 18 μM (FIG. 2B).

55

15

These results show that the COI1/JAZ complex, rather than COI1 alone, functions as the genuine high-affinity jasmonate receptor in a co-receptor form.

#### Example 2

## Characterization of JAZ1 Peptides

The COI1-binding region of the JAZ proteins has previously been mapped to the carboxy-terminal Jas motif, which is characterized by the SLXXFXXKRXXRXXXXXPY consensus sequence (SEQ ID NO:30) preceded by two consecutive basic residues. As shown in Example 1, mutation of the conserved phenylalanine residue to alanine is sufficient to abolish the high affinity interaction between coronatine and COI1/JAZ6 (FIG. 1A).

Previous studies have shown that the highly conserved PY sequence at the C-terminal end of the Jas motif plays a role in JAZ localization and stability in vivo, but that the sequence was not necessary for ligand-dependent COI1-JAZ interaction. This was confirmed by an experiment showing that truncation of the PY motif in JAZ1 had little effect on in vitro ligand binding activity (FIG. 3). To further map the minimal region of the Jas motif required for high affinity ligand binding with COI1, the recombinant JAZ1 protein was replaced with a set of synthetic JAZ1 peptides in a ligand binding assay.

The JAZ1 degron of SEQ ID NO:1 (R205-Y226), which <sup>30</sup> spans the central conserved Jas motif plus the two additional amino-terminal basic residues, did not bind to COI1 with high affinity in the presence of coronatine (FIG. 4). These results indicate that residues N-terminal to Arg205 must participate in the COI1/JAZ interaction. Various derivatives of the JAZ1 <sup>35</sup> degron sequence, as well as derivatives of the JAZ6 and JAZ7 degron sequences, were analyzed for their ability to bind COI1 with high affinity in the presence of coronatine. These derivatives are set forth in Table 1.

TABLE 1

Peptide	SEQ ID NO	Sequence
JAZ1 Jas motif		PIARRASLHRFLEKRKDRVTSK APY
JAZ1 degron	1	RRASLHRFLEKRKDRVTSKAPY
JAZ1 + 1 extension	2	ARRASLHRFLEKRKDRV
JAZ1 + 2 extension	3	IARRASLHRFLEKRKDRV
JAZ1 + 3 extension	4	PIARRASLHRFLEKRKDRV
JAZ1 + 4 extension	5	LPIARRASLHRFLEKRKDRV
JAZ1 + 5 extension	6	ELPIARRASLHRFLEKRKDRV
JAZ1 + 5 extension + PY motif	7	ELPIARRASLHRFLEKRKDRVT SKAPY
JAZ1 + polyA extension	8	AAAAARRASLHRFLEKRKDRV
JAZ1 + 5 extension from JAZ6	9	VERIARRASLHRFLEKRKDRV
JAZ6 + 5 extension	10	VERIARRASLHRFFAKRKDRV

16

TABLE	1-continued

Peptide	SEQ ID NO	Sequence
JAZ6 + 10 extension	11	QQHQVVERIARRASLHRFFAKR KDRV
JAZ7 + 5 extension	12	YQKASMKRSLHSFLQKRSLRI

The JAZ1 +1, +2, +3, +4, and +5 extensions (SEQ ID NOs:2-6, respectively) added on 1, 2, 3, 4, and 5 residues, respectively, to the N-terminus of JAZ1 while removing six residues from the C-terminus. The added residues were derived from the residues that are normally present N-terminal to the degron in JAZ1. An additional JAZ1 peptide (SEQ ID NO:7) contained the same additional N-terminal amino acids as the +5 extension but with the six C-terminal residues added back on.

The JAZ1 +4 and +5 extensions (SEQ ID NOs:5 and 6, respectively) were found to bind COI1 in the presence of coronatine with a much higher affinity than the JAZ1 degron and the +1, +2, and +3 extensions, with the +5 extension exhibiting the highest degree of binding (FIG. 4). The JAZ1 +5 extension peptide was found to permit coronatine binding with a  $K_D$  (~108 nM) comparable to that of the full-length JAZ1 protein (FIG. 5). The JAZ1 +5 extension with the six C-terminal residues added back on also exhibited significant binding (FIG. 6, "JAZ1-extension +5+PY").

To test the specificity of the JAZ1 peptide tags of SEQ ID NOs:5, 6, and 7, an additional set of JAZ peptide tags was developed. The first two were essentially identical to the JAZ1 +5 extension in that they contained residues 1-16 of the JAZ1 degron plus a five amino acid N-terminal extension. However, the sequence of the extension was different. The first of these (SEQ ID NO:8) utilized a polyalanine extension, while the second (SEQ ID NO:9) utilized a five amino acid extension derived from JAZ6. The remaining JAZ peptide tags were based on the degrons of JAZ6 and JAZ7. These included two JAZ6 peptides that contained the JAZ6 degron plus an additional five and ten N-terminal residues, respectively (SEQ ID NOs: 10 and 11, respectively) and one JAZ7 peptide that contained the JAZ7 degron plus an additional five N-terminal residues (SEQ ID NO: 12). None of the additional - 45 JAZ peptide tags exhibited significant binding to COI1 in the presence of coronatine (FIGS. 4 and 6). These results suggest that the system disclosed herein maintains a great deal of selectivity for side chain chemistry, and that simply inserting five "filler" amino acid residues at the N-terminus of the 50 peptide tag is insufficient to promote COI1 binding.

## Example 3

## Structural Relationship of COI1/JAZ1 and Coronatine

To evaluate the structure mechanism by which COI1/JAZ1 co-receptor senses jasmonate, crystal structures were obtained for COI1/ASK1/JAZ1 peptides complexed with either coronatine or (3R,7S)-JA-Ile.

Crystals were grown at 4° C. by the hanging-drop vapor diffusion method with 1.5 μL protein complex samples containing COI1/ASK1, JAZ1 peptide, and hormone compound at 1:1:1 molar ratio mixed with an equal volume of reservoir solution containing 100 mM BTP, 1.7-1.9 M ammonium phosphate, and 100 mM NaCl, pH 7.0. Diffraction quality crystals were obtained by the micro-seeding method at 4° C.

17

The crystals all contained eight copies of the complex in the asymmetric unit. The data sets were collected at the Advanced Light Source in Lawrence Berkeley National Laboratory as well as the GM/CA-CAT 23 ID-B beamline at the Advanced Photon Source in Argonne National Laboratory using crystals flash-frozen in the crystallization buffers supplemented with 15-20% ethylene glycol at  $-170^{\circ}$  C. Reflection data were indexed, integrated, and scaled with the HKL2000 package. All crystal structures were solved by molecular replacement using the program Phaser and the TIR1/ASK1 structure as search model. The structural models were manually built in the program O and refined using CNS and PHENIX. All final models had 96-98% of residues in the favored region and 0% in disallowed region of the Ramachandran plot.

18

Without the JAZ degron peptide bound, the keto group of the ligand is accessible to solvent (FIG. 10). The rest of the cyclopentanone ring of both JA-Ile and coronatine is sandwiched between the aromatic groups of F89 and Y444 of COI1, stabilized by hydrophobic packing. The cyclohexene ring of coronatine provides a rigid surface area for close packing with F89, whereas the more flexible and extended pentenyl side chain of JA-Ile is more loosely accommodated by a hydrophobic pocket formed by A86, F89 and L91 from loop 2 as well as L469 and W519 from the LRRs (FIG. 11A). Differences at this interface probably explain the approximately tenfold higher affinity of coronatine over (3R,7S)-JA-Ile detected in binding assays. Deeper in the ligand-binding pocket, the common amide and carboxyl groups of JA-Ile and

TABLE 2

		COI1/ASK1/JAZ1 degron/coronatine	COI1/ASK1/JAZ1 +5 extension/ coronatine	COI1/ASK1/JAZ1 +5 extension/JA-Ile
Data colle	ection			
Space gr Cell dimensions	roup a, b, c (Å)	P21 121.8, 221.5, 148.5	P21 123.2, 220.8, 149.5	P21 122.3, 220.8, 148.7
difficusions	$\alpha, \beta, \gamma$ (°)	90.0, 104.5, 90.0	90.0, 104.5, 90.0	90.0, 104.5, 90.0
Resolutio R <sub>sym</sub> //o/	n (Å)	2.80 (2.80-2.90) 0.103 (0.816) 16.7 (2.0)	3.35 (3.35-3.41) 0.119 (0.700) 14.0 (2.1)	3.18 (3.18-3.31) 0.088 (0.462) 17.2 (2.8)
Completeness (%) Redundancy Refinement		100 (100) 3.9 (3.8)	92.9 (94.6) 3.6 (3.3)	97.0 (93.3) 3.1 (2.7)
Resolutio No. reflec R <sub>work</sub> /R	tions	50-2.80 174,966 0.235/0.270	50-3.35 95,997 0.225/0.270	50-3.18 116,337 0.223/0.264
R.m.s deviations	Bond lengths (Å)	0.008	0.010	0.010
	Bond angles (°)	1.676	1.271	1.556

The crystal structure of COI1 revealed a TIR1-like overall architecture, with the N-terminal tri-helical F-box motif bound to ASK1 and a C-terminal horseshoe-shaped solenoid domain formed by 18 tandem leucine-rich repeats (FIGS. 7A and 7B). Similar to TIR1, the top surface of the COI1 leucine-rich repeat (LRR) domain has three long intra-repeat loops (loops 2, 12, and 14) that are involved in hormone and polypeptide substrate binding. Unlike TIR1, however, a fourth long loop (loop C) in the C-terminal capping sequence

of the COI1 LRR domain folds over loop 2, partially covering

it from above (FIGS. 7B and 7C).

Despite their similar overall fold, crystal structure analysis revealed that COI1 has evolved a hormone binding site that is distinct from that of TIR1. Configured between loop 2 and the 55 inner wall of the LRR, the ligand binding pocket of COI1 is

exclusively encircled by amino acid side chains (FIG. 8). Many of the pocket-forming residues on COI1 are large in size and carry a polar head group (FIG. 9). These properties allow them to be mold a binding pocket into a specific shape while forming close interactions with each chemical moiety of the ligand. These close interactions are critical to proper hormone sensing of the complex. In the binding pocket, both JA-Ile and coronatine sit in an 'upright' position with the keto group of their common cyclopentanone ring pointing up and 65

forming a triangular hydrogen bond network with R496 and

Y444 of COI1 at the pocket entrance (FIG. 8).

coronatine bind to the bottom of the binding site by forming a salt bridge and hydrogen bond network with three basic residues of COI1:R85, R348 and R409 (FIGS. 8A and 8B). Together, these arginine residues constitute the charged floor of the ligand pocket. Y386 reinforces the interactions from above by forming a hydrogen bond with the amine group of the ligand. In doing so, Y386 approaches the cyclopentanone ring of the ligand, narrowing the pocket entrance and creating a hydrophobic cave below. The rest of the basin is carved out by V411, A384 and the aliphatic side chain of Arg 409 (FIG. 11B). The ethyl-cyclopropane group of coronatine and the isoleucine side chain of JA-Ile can both comfortably fit in this space due to their similar size and hydrophobicity. The nature of the cave explains the preference of COI1 for jasmonate conjugates containing a moderately sized hydrophobic amino acid. Although most of the ligand is buried inside the binding site, the keto group at the top and the carboxyl group at the bottom remain exposed, available for additional interactions with the JAZ portion of the co-receptor (FIG. 10).

The JAZ1 degron peptide adopts a bipartite structure with a loop region followed by an  $\alpha$ -helix to assemble with the COI1/JA complex. The hallmark of the JAZ1 degron is the N-terminal five amino acids identified in the radioligand binding assay. In a largely extended conformation, this short sequence lies on top of the hormone-binding pocket and simultaneously interacts with both COI1 and the ligand,

effectively trapping the ligand in the pocket (FIGS. 12A and 12B). At the N-terminal end, L201 of the JAZ1 peptide is embedded in a hydrophobic cavity presented by surface loops on top of COI1 (FIG. 13A).

At the C-terminal end, A204 of JAZ1 uses its short side chain to pack against the keto group of the ligand and F89 of COI1 (FIGS. 11A and 13A). The same alanine residue of JAZ1 also donates a hydrogen bond through its backbone amide group to the keto moiety of the ligand emerging from the pocket (FIG. 13A). The middle region of the five-aminoacid sequence is secured to the COI1 jasmonate complex through a hydrogen bond formed between the backbone carbonyl of P202 in JAZ1 and the ligand-interacting COI1 residue R496, which is critical for the hormone-dependent COI1/ JAZ interaction (data not shown). In agreement with its important role in forming the JA-Ile co-receptor, this short N-terminal region of the JAZ degron completely covers the opening of the ligand-binding pocket, conferring high-affinity binding to the hormone. The close interaction between the 20 hormone and the co-receptor complex provides a plausible structural explanation for the favorable binding of the (3R, 7S)-JA-Ile isomer, as the stereochemistry at the 7 position of (3R,7R)-JA-Ile may place the aliphatic chain unfavorably close to nearby JAZ1 and COI1 residues (FIG. 11A).

Within the JAZ1 degron, two conserved basic residues, R205 and R206, were previously shown to have an important role in hormone-induced COI1 binding. In the structure, R205 contributes to COI1 binding by directly interacting with loop 12, whereas R206 points in the opposite direction and inserts deeply into the central tunnel of the COI1 solenoid. Approaching the bottom of the ligand-binding pocket, the guanidinium group of the R206 side chain joins the three basic COI1 residues that form the pocket floor and interacts directly with the carboxyl group of the ligand (FIG. 13B). Thus, the N-terminal seven amino acids (ELPIARR) of the JAZ1 degron peptide act as a clamp that wraps the ligand-binding pocket from top to bottom, closing it completely (FIG. 12B).

The highly conserved C-terminal half of the JAZ1 degron forms an amphipathic  $\alpha$ -helix that strengthens the JAZ1/COI1 interaction by binding to the top surface of the COI1 LRR domain, adjacent to the ligand-binding site (FIG. 12A). With its N-terminal end directly packing against loop 2 of 45 COI1, the Jas motif helix blocks the central tunnel of the COI1 LRR solenoid like a plug. The N-terminal half of the Jas motif helix is characterized by three hydrophobic residues (L209, F212 and L213) which are aligned on the same side of the helix and form a hydrophobic interface with COI1 (FIG. 50 13C).

By soaking the COI1-ASK1 crystals with coronatine and a sufficiently high concentration of JAZ1 degron peptide lacking the N-terminal ELPIA sequence, the complex formed by COI1, coronatine, and the isolated Jas motif helix was trapped in the crystal (Table 2). This indicates that the  $\alpha$ -helix may provide a low-affinity anchor for docking the JAZ protein on COI1.

## Example 4

#### Identification of COI1 Cofactor

The crystal structure of TIR1 revealed an unexpected inositol hexakisphosphate (InsP<sub>6</sub>) molecule bound in the centre of 65 the protein underneath the auxin-binding pocket. Sequence homology between COI1 and TIR1 suggests that COI1 might

20

contain a similar small molecule. Before crystallization, the recombinant COI1/ASK1 complex was analyzed by structural mass spectrometry.

Nano-electrospray ionization mass spectrometry (MS) and tandem MS (MS/MS) experiments were performed on a SynaptHDMS instrument. Before MS analysis, 50 µL of a 16 mg ml<sup>-1</sup> solution of COI1/ASK1 in 20 mM Tris-HCl, pH 8, 0.2 M NaCl, and 5 mMDTT was buffer-exchanged twice into 0.5M ammonium acetate solution using Bio-Rad Biospin columns. To improve desolvation during ionization, samples were diluted 1:4 in 0.5 M ammonium acetate, and isopropanol was added to a final concentration of 5%. Typically an aliquot of 2 mL solution was loaded for sampling via nano-ESI capillaries, which were prepared in-house from borosilicate glass tubes as described previously (Nettleton 1998). The conditions within the mass spectrometer were adjusted to preserve non-covalent interactions. The following experimental parameters were used: capillary voltage up to 1.26 kV, sampling cone voltage 150 V, and extraction cone voltage 6 V, MCP 1590. For tandem MS experiments peaks centered at m/z 4,564 and 4,588 were selected in the quadrupole and collision energy up to 65 V was used. Argon was used as a collision gas at maximum pressure. All spectra were calibrated externally using a solution of cesium iodide (100 mg 25 ml<sup>-1</sup>). Spectra are shown with minimal smoothing and without background subtraction.

Nano-electrospray mass spectra of the intact COI1/ASK1 complex revealed two populations differing by a mass of ~568 Da, indicating that a small molecule was indeed copurified with the proteins (FIGS. 15 and 16). As shown in FIG. 16, only one population of the complex corresponding to COI/ASK1 was apparent at both at low and high collision energy. At high collision energy, the COI/ASK1 complex dissociates into its different subunits and one population of COI1 appears in the spectrum. This population of COI1 has a calculated mass of 67,944±1 Da, which is in agreement with the theoretical mass of COI1 (67,947 Da). At low collision energy, the COI1/ASK1/ligand complex is apparent. However, elevating the collision energy releases some of the bound ligand and results in the appearance of a stripped COI1/ASK1 complex. The theoretical mass of the apo COI1/ ASK1 complex is 86,458 Da, which is in close agreement with the observed mass of 86,543±28 Da. The mass of the COI1/ASK1/ligand complex was found to be 87,112±15 Da, suggesting that the mass of the ligand is around 568±28 Da. The fact that both masses carry a charge of +19 indicates a neutral loss of the ligand, meaning that it cannot be detected in the spectrum. At high collision energy, some of the complex dissociates into its different subunits and two populations of COI1 appear in the spectrum. The smaller form, with a calculated mass of 67,952±5 Da, fits the theoretical mass of COI1 (67,946.5 Da), whereas the other population, with a calculated mass of 68,518±4 Da, corresponding to COI1ligand, suggest that the mass of the ligand is around 568±5

The mass-spectrometry-derived molecular mass of the unknown compound is different from the mass of  $nsP_6$  (651 Da) but matches that of an inositol pentakisphosphate ( $InsP_5$ ) molecule. Unfortunately, mass spectrometry analyses of either the native COI1/ASK1 complex or the denatured proteins were unable to achieve direct mass analysis of the small molecule.

To investigate the identity of the unknown compound, it was first estimated that the molecule contains four or five phosphate groups by <sup>31</sup>P nuclear magnetic resonance (NMR) of trypsin-digested COI1/ASK1 complex (data not shown). To identify unequivocally the unknown molecule, steps were

taken to purify it away from the COI1/ASK1 complex in a quantity sufficient for <sup>1</sup>H NMR analysis. The high phosphate content of the molecule allowed us to trace it through a multi-step purification procedure (FIG. 17). Phenol was melted at 68° C. and equilibrated with equal parts 0.5 M 5 Tris-HCl, pH 8.0 until a pH of 7.8 was reached. The equilibrated phenol was then topped with 0.1 volume 100 mM Tris-HCl, pH 8.0 and stored at 4° C. For extraction, 30-40 mg of 1 mg ml<sup>-1</sup> COI1/ASK1 protein was mixed in small batches with equal parts equilibrated phenol at room temperature. The 10 samples were inverted and incubated for 30 minutes until phase separation occurred. With 30 second vortexing, the samples were incubated at room temperature for 30 minutes and spun at 15,000 rpm for 5 minutes. The aqueous phase was removed as a primary extraction. Equal parts of a solution 15 containing 25 mM Tris-HCl, pH 8.0 was added to the phenol and collected as above as a secondary extraction. The primary and secondary extractions were combined and diluted 10× in 25 mM Tris-HCl, pH 8.0, then further purified by gravity flow on O sepharose high-performance anion exchange resin (GE 20 Healthcare). Following column wash with 10x column volumes of 0.1 N formic acid, stepwise elution was performed with 23 column volumes of 0.1 N formic acid (Thermo Scientific) with increasing concentrations of ammonium formate (Sigma) from 0 to 2 M. Fractions were analyzed for phos- 25 phate content by the wet-ashing method with perchloric acid in Pyrex culture tubes (13×100 mm). Typically, samples of 50-100 µL were ashed with 100-200 mL 70% perchloric acid (purified by redistillation, Sigma). Ashing was performed by heating the sample over a Bunsen-type burner with continuous shaking to prevent bumping. When the sample stopped emitting white smoke, the reaction was considered complete and then heated to dryness. 500  $\mu L$  of distilled water was added to the room temperature tubes and vortexed. 100 µL samples containing up to 10 nmol inorganic phosphate were 35 assayed for phosphate by a modification of a published procedure (Sadrzadeh 1993). A total of 125 µL of acid molybdate color reagent was added and the samples were incubated and covered at room temperature for 12-14 hours (overnight) for full color development (total volume 225 µL). Plates were 40 read at 650 nm and unknowns were determined from the linear regression of the standard curve (0-10 nmol NaH<sub>2</sub>PO<sub>4</sub> per well). All assays were done in triplicate. Final fractions containing phosphate were combined and lyophilized repeatedly to remove residual ammonium formate.

After isolation of 150 nmol of the purified small molecule, a series of one-dimensional and two-dimensional NMR data were acquired, including a highly informative homonuclear total correlation (TOCSY) spectrum. NMR spectra were acquired on a Varian INOVA600 spectrometer equipped with 50 a cold probe using 200  $\mu$ M samples of purified compound X or synthetic inositol-1,2,4,5,6-pentakisphosphate (Cayman Chemical) dissolved in D<sub>2</sub>O. TOCSY spectra were acquired with mixing times of 35 or 50 ms, processed with NMRPipe and visualized with NMRView.

The observed chemical shifts and TOCSY cross-peak patterns are clearly characteristic of inositol phosphates (FIG. **18**). A comparison with previously reported NMR spectra of various inositol phosphates established that the unknown compound is either D- or L-inositol-1,2,4,5,6-pentakisphosphate (Ins(1,2,4,5,6) $P_5$ ; FIG. **18**). This conclusion was further supported by the TOCSY spectrum of synthetic Ins(1,2,4,5,6) $P_5$  (FIG. **19**) and the subsequently acquired negative ion electrospray ionization mass spectrometry spectrum of the compound (FIG. **20**).

As shown in FIG. 20, the negative-ion ESI-MS spectrum of the unknown contained the major ion at m/z 192.3

 $((579.8951-3\times1.0078)/3)$ , corresponding to the [M-3H]<sup>3-</sup> ion of inositol pentakisphosphate (InsP<sub>5</sub>), and the ion at m/z 288.9  $((579.8951-2\times1.0078)/2)$ , corresponding to the  $[M-2H]^{2-}$  ion of InsP<sub>5</sub>. The  $[M-H]^{-}$  ion expected at m/z 579.9 was absent. The ions seen at m/z 199.7 and 207.1 correspond to the sodiated ions of InsP<sub>5</sub> seen as the [M+Na-4H]<sup>3-</sup>, and [M+2Na-5H]<sup>3</sup> ions, respectively; and the ions at m/z 299.9 and 311.9 correspond to the  $[M+Na-3H]^{2-}$  and [M+2Na-4H]<sup>2-</sup> ions, respectively. The spectrum also contains ions at m/z 499 ([M-H—HPO<sub>3</sub>]<sup>-</sup>), 419 ([M-H-2HPO<sub>3</sub>]<sup>-</sup>), and 441 ([M+Na-2H-2HPO<sub>3</sub>]<sup>-</sup>), arising from various losses of the phosphate residues of the molecule. The presence of the ion at m/z 499 (579.9-HPO<sub>3</sub>) is consistent with the observation of the ions at m/z 249 ([M-2H- $HPO_3]^{-2}$ , 259.9 ([M+Na-3H—HPO<sub>3</sub>]<sup>-2</sup>), and 165.7 ([M-3H—HPO<sub>3</sub>]<sup>-3</sup>), representing the various deprotonated InsP<sub>4</sub> seen as doubly and triply charged anions. The ion at m/z 419 represents a deprotonated InsP3 arising from loss of two HPO<sub>3</sub> residues; while the ion at m/z 441 represents a monosodiated InsP<sub>3</sub> anion. The presence of the ions at m/z 419 and 441 is also consistent with the observation of the doubly charged ions at m/z 209 and 219, corresponding to the  $[M-2H-2HPO_3]^{-2}$  and  $[M+Na-3H-2HPO_3]^{-2}$  ions, respectively. The assignments of the ions observed are listed in Table 3. These ions were also observed for  $Ins(1,2,3,4,5)P_5$ and Ins(1,2,4,5,6)P<sub>5</sub> standards when subjected to ESI under the same condition, indicating that the unknown compound is an InsP<sub>5</sub>. This InsP<sub>5</sub> structure is further confirmed by the MSn (n=2,3,4,5) mass spectra of the  $[M-3H]^{3-}$  ion at m/z 192.3 and of the [M-2H]<sup>2-</sup> ion at m/z 288.9 deriving from the unknown compound and from the  $Ins(1,2,3,4,5)P_5$  and  $Ins(1,2,4,5,6)P_5$ standards.

TABLE 3

m/z	Structure
III/Z	Structure
499	[M – H – HPO <sub>3</sub> ] <sup>–</sup>
441	$[M + Na - 2H - 2HPO_3]^-$
419	$[M - H - 2HPO_3]^-$
311	$[M + 2Na - 4H]^{-2}$
300	$[M + Na - 3H]^{-2}$
289	$[M - 2H]^{-2}$
271	$[M + 2Na - 2H - 2HPO_3]^{-2}$
268	$[M + K - 3H - 2HPO_3]^{-2}$
259.9	$[M + Na - 3H - HPO_3]^{-2}$
249	$[M - 2H - HPO_3]^{-2}$
219	$[M + Na - 3H - 2HPO_3]^{-2}$
212	$[M + Na + K - 5H]^{-3}$
209	$[M - 2H - 2HPO_3]^{-2}$
207	$[M + 2Na - 5H]^{-3}$
203	$[M + Na - 4H]^{-3}$
199.7	$[M + Na - 4H]^{-3}$
192.3	$[M - 3H]^{-3}$
165.7	$[M - 3H - HPO_3]^{-3}$
97	$H_2PO_4^-$
79	PO <sub>3</sub> -

Consistent with the binding of a small molecule cofactor, the crystal structure of COI1 (Example 3) showed strong unexplained electron densities clustered in the middle of the COI1 LRR domain. Like InsP<sub>6</sub> in TIR1, these extra densities in COI1 are located directly adjacent to the bottom of the ligand binding pocket of the jasmonate co-receptor, interacting with multiple positively charged COI1 residues (FIG. 21A). Unexpectedly, these islands of electron density cannot be explained by an Ins(1,2,4,5,6)P<sub>5</sub> molecule. Instead, their intensity, overall symmetry, and poor connectivity indicate that they belong to multiple free phosphate molecules. Because a high concentration of ammonium phosphate was

used as the major precipitant for crystallizing the JA correceptor, it was postulated that the InsP<sub>5</sub> molecule that copurified with COI1 was later displaced by phosphate molecules in the crystallization drops. In support of this scenario, the concave surface of the COI1 solenoid fold surrounding the phosphates is highly basic and decorated with residues conserved in plant COI1 orthologs, indicating a functionally important surface area (FIGS. 9, 21B, 22).

The highly selective co-purification of two different inositol phosphates, InsP<sub>5</sub> and InsP<sub>5</sub>, with two homologous plant hormone receptors, COI1 and TIR1, implies that the proper function of the two F-box proteins might require the binding of specific inositol phosphates. To assess the functional role of Ins(1,2,4,5,6)P<sub>5</sub> in the COI1/JAZ1 co-receptor, a protocol was developed for stripping the co-purified InsP<sub>5</sub> from COI1 without denaturing the protein. Briefly, proteins were mixed with 10% glycerol and incubated in 2 M ammonium phosphate, 100 mM Bis-Tris propane, pH 7.0, 200 mM NaCl, and 10% glycerol at 4° C. for >24 hours with a minimum of  $3\times 20$ buffer changes at 100× sample volume. Samples were then transferred to 20 mM Tris-HCl, pH 8.0, 200 mM NaCl, and 10% glycerol at 4° C. for >24 hours with a minimum of three buffer changes at 100× sample volume. Inositol phosphate rescue experiments were conducted according to the radioli- 25 gand binding assays described above in the presence of 300 nM <sup>3</sup>H-coronatine with nonspecific binding determined in the presence of 300 µM coronatine.

The resulting COI1/ASK1 complex was tested in a ligand-binding based reconstitution assay. As shown in FIG. **23**A, 30 untreated COI1 formed a high-affinity jasmonate co-receptor with JAZ1. Addition of exogenous  $Ins(1,2,4,5,6)P_5$  did not significantly change its activity. In contrast, the dialyzed COI1 sample completely lacked ligand binding by itself and showed only trace activity in the presence of JAZ1. Supplementation with either synthetic  $Ins(1,2,4,5,6)P_5$  (FIG. **23**B) or the purified and NMR analyzed  $InsP_5$  sample (data not shown) rescued the interaction in a dose-dependent manner and with a half-maximum effective concentration (EC<sub>50</sub>) of 27 nM (FIG. **23**C). From this reconstitution result, it was 40 concluded that  $Ins(1,2,4,5,6)P_5$  binding is crucial for the jasmonate coreceptor to perceive the hormone with high sensitivity.

A close examination of the phosphate molecules in the available COI1 structure indicates a mechanism by which the 45 inositol phosphate molecule may modulate the activity of the jasmonate co-receptor. Among four COI1-bound phosphates, one stands out by binding at a critical position in the jasmonate co-receptor. This phosphate molecule interacts simultaneously with four basic residues at the bottom of the 50 ligand-binding pocket, namely Arg 206 in the JAZ1 degron and the three COI1 arginine residues that form the floor of the pocket. As a result, a tetragonal bipyramidal interaction network is formed among four molecules at the core of the jasmonate co-receptor assembly. The four arginines from 55 COI1 and JAZ1 sit at the four corners of the central plane, interacting with the hormone above and the phosphate below (FIG. 24).

As the free phosphate molecule probably mimics the action of a phosphate group on  $InsP_5$ , this four-molecule junction, 60 together with additional phosphate-COI1 interactions seen in the crystal, conceivably represents the structural basis for  $InsP_5$  potentiation of the jasmonate coreceptor. Consistent with this interpretation, coronatine-induced formation of a COI1/JAZ1 complex was readily abolished by mutation of 65 select COI1 residues adjacent to the phosphates, but not in contact with the hormone (FIG. **25**).

24

The reconstitution assay was used to further investigate the specificity of jasmonate co-receptor regulation by inositol phosphates (FIG. 14A). Notably, inositol-1,4,5,6-tetrakisphosphate supports the activity of the COI1/JAZ1 co-receptor, whereas the second messenger signaling molecule inositol-1,4,5-trisphosphate does not. Addition of a phosphate to InsP<sub>5</sub>, which gives rise to InsP<sub>6</sub>, is also not favorable for activity. Although saturation binding of <sup>3</sup>H-coronatine is stimulated by both Ins(1,2,4,5,6)P<sub>5</sub> and InsP<sub>6</sub> with similar 1K<sub>d</sub> values (30 nM and 37 nM, respectively), the two inositol phosphates yield markedly different  $\mathbf{B}_{max}$  values for coronatine binding, indicating that InsP6 is significantly less efficacious in activating the co-receptor despite having equal affinity as Ins(1,2,4,5,6)P<sub>5</sub> (FIG. 14B). Functional selectivity of COI1 for the inositol phosphate cofactor is consistent with the conservation of the putative inositol-phosphate-binding site, which is distinct in amino acid sequence from the InsP<sub>6</sub>binding site in TIR120 (FIG. 9).

#### Example 5

### Targeted Degradation of a Target Protein

Green fluorescent protein (GFP) will be tagged with the JAZ1 +5 extension peptide tag of SEQ ID NO:6 in budding yeast cells (e.g., *Saccharomyces cerevisiae*) and/or mammalian cells. Where budding yeast cells are used, the tagged protein construct will be cloned into a standard yeast shuttling plasmid under the control of a strong, stable promoter, and the plasmid will be stably inserted into the yeast genome via chromosomal recombination sequences using methods well known in the art. Where mammalian cells are used, the gene encoding the tagged protein construct will be introduced via transient transfection or stable cell line generation.

The cells will be further engineered to express *Arabidopsis* COI1 or a homolog thereof under the control of an inducible promoter. For example, exogenous COI1 expression may be placed under the control of a galactose promoter, such that expression may be controlled by sugar ratio.

After stable GFP signal has been monitored qualitatively using standard techniques such as microscopy and/or quantitatively using techniques such as standard plate readers and/or flow cytometry methods, expression of *Arabidopsis* COI1 or a homolog thereof will be induced. COI1 expression should not significantly affect GFP signal levels.

Cells will be treated with titrating levels of coronatine. Cells will be harvested and fixed at various timepoints, and GFP signal will be quantified to determine the rate of GFP degradation. Degradation will increase as coronatine levels increase.

Additional experiments may be performed using one or more of the other JAZ1 peptide tags disclosed herein to determine the efficacy of slight changes to the peptide sequence. Similarly, additional experiments may be performed using molecules other than coronatine that bind to the COI1/JA-Ile binding pocket of COI1.

As stated above, the foregoing is merely intended to illustrate various embodiments of the present invention. The specific modifications discussed above are not to be construed as limitations on the scope of the invention. It will be apparent to one skilled in the art that various equivalents, changes, and modifications may be made without departing from the scope of the invention, and it is understood that such equivalent embodiments are to be included herein. All references cited herein are incorporated by reference as if fully set forth herein.

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Arg Glu Met Glu His Pro Ala His Ile Leu Ala Tyr Tyr Ser Leu Ala 570 Gly Gln Arg Thr Asp Cys Pro Thr Thr Val Ile Val Leu Arg Glu Pro 585 Met <210> SEQ ID NO 17 <211> LENGTH: 595 <212> TYPE: PRT <213 > ORGANISM: Oryza sativa <400> SEQUENCE: 17 Met Gly Glu Val Pro Glu Pro Arg Arg Leu Asn Arg Ala Leu Ser Phe Asp Asp Trp Val Pro Asp Glu Ala Leu His Leu Val Met Gly His 20 25 30Val Glu Asp Pro Arg Asp Arg Glu Ala Ala Ser Arg Val Cys Arg Arg Trp His Arg Ile Asp Ala Leu Thr Arg Lys His Val Thr Val Ala Phe Cys Tyr Ala Ala Arg Pro Ala Arg Leu Arg Glu Arg Phe Pro Arg Leu 65 70 75 80 Glu Ser Leu Ser Leu Lys Gly Lys Pro Arg Ala Ala Met Tyr Gly Leu Ile Pro Asp Asp Trp Gly Ala Tyr Ala Ala Pro Trp Ile Asp Glu Leu 105 Ala Ala Pro Leu Glu Cys Leu Lys Ala Leu His Leu Arg Arg Met Thr 120 Val Thr Asp Ala Asp Ile Ala Ala Leu Val Arg Ala Arg Gly His Met 135 Leu Gln Glu Leu Lys Leu Asp Lys Cys Ile Gly Phe Ser Thr Asp Ala Leu Arg Leu Val Ala Arg Ser Cys Arg Ser Leu Arg Thr Leu Phe Leu 170 Glu Glu Cys His Ile Thr Asp Lys Gly Gly Glu Trp Leu His Glu Leu Ala Val Asn Asn Ser Val Leu Val Thr Leu Asn Phe Tyr Met Thr Glu Leu Lys Val Ala Pro Ala Asp Leu Glu Leu Leu Ala Lys Asn Cys Lys Ser Leu Ile Ser Leu Lys Met Ser Glu Cys Asp Leu Ser Asp Leu Ile Ser Phe Phe Gln Thr Ala Asn Ala Leu Gln Asp Phe Ala Gly Gly Ala 250 Phe Tyr Glu Val Gly Glu Leu Thr Lys Tyr Glu Lys Val Lys Phe Pro Pro Arg Leu Cys Phe Leu Gly Leu Thr Tyr Met Gly Thr Asn Glu Met 280 Pro Val Ile Phe Pro Phe Ser Met Lys Leu Lys Lys Leu Asp Leu Gln 295 Tyr Thr Phe Leu Thr Thr Glu Asp His Cys Gln Ile Ile Ala Lys Cys 310 315

Pro Asn Leu Leu Ile Leu Glu Val Arg Asn Val Ile Gly Asp Arg Gly

Leu Glu Val Val Gly Asp Thr Cys Lys Lys Leu Arg Arg Leu Arg Ile

Glu Arg Gly Asp Asp Pro Gly Leu Gln Glu Glu Gln Gly Val Ser Gln Leu Gly Leu Thr Ala Val Ala Val Gly Cys Arg Glu Leu Glu Tyr Ile Ala Ala Tyr Val Ser Asp Ile Thr Asn Gly Ala Leu Glu Ser Ile Gly Thr Phe Cys Lys Asn Leu Tyr Asp Phe Arg Leu Val Leu Leu Asp Arg Glu Arg Gln Val Thr Asp Leu Pro Leu Asp Asn Gly Val Cys \$420\$ \$425\$ \$430Ala Leu Leu Arg Asn Cys Thr Lys Leu Arg Arg Phe Ala Leu Tyr Leu Ser Gly Asn Ile Gln Tyr Met Leu Leu Gly Asn Val Gly Glu Ser Asp 470 His Gly Leu Ile Arg Phe Ala Val Gly Cys Thr Asn Leu Gln Lys Leu Glu Leu Arg Ser Cys Cys Phe Ser Glu Arg Ala Leu Ser Leu Ala Val Leu Gln Met Pro Ser Leu Arg Tyr Ile Trp Val Gln Gly Tyr Arg Ala 520 Ser Gln Thr Gly Leu Asp Leu Leu Leu Met Ala Arg Pro Phe Trp Asn 535 Ile Glu Phe Thr Pro Pro Ser Pro Glu Ser Phe Asn His Met Thr Glu Asp Gly Glu Pro Cys Val Asp Ser His Ala Gln Val Leu Ala Tyr Tyr 570 Ser Leu Ala Gly Arg Arg Ser Asp Cys Pro Gln Trp Val Ile Pro Leu His Pro Ala 595 <210> SEQ ID NO 18 <211> LENGTH: 603 <212> TYPE: PRT <213 > ORGANISM: Solanum lycopersicum <400> SEQUENCE: 18 Met Glu Glu Arg Asn Ser Thr Arg Leu Ser Ser Ser Thr Asn Asp Thr Val Trp Glu Cys Val Ile Pro Tyr Ile Gln Glu Ser Arg Asp Arg Asp Ala Val Ser Leu Val Cys Lys Arg Trp Trp Gln Ile Asp Ala Ile Thr Arg Lys His Ile Thr Met Ala Leu Cys Tyr Thr Ala Lys Pro Glu Gln 55 Leu Ser Arg Arg Phe Pro His Leu Glu Ser Val Lys Leu Lys Gly Lys Pro Arg Ala Ala Met Phe Asn Leu Ile Pro Glu Asp Trp Gly Gly Tyr Val Thr Pro Trp Val Met Glu Ile Thr Lys Ser Phe Ser Lys Leu Lys

	Concinaca														
			100					105					110		
Ala	Leu	His 115	Phe	Arg	Arg	Met	Ile 120	Val	Arg	Asp	Ser	Asp 125	Leu	Glu	Leu
Leu	Ala 130	Asn	Arg	Arg	Gly	Arg 135	Val	Leu	Gln	Val	Leu 140	Lys	Leu	Asp	Lys
Cys 145	Ser	Gly	Phe	Ser	Thr 150	Asp	Gly	Leu	Leu	His 155	Ile	Ser	Arg	Ser	Сув 160
ГÀа	Asn	Leu	Arg	Thr 165	Leu	Leu	Met	Glu	Glu 170	Ser	Tyr	Ile	Ile	Glu 175	Lys
Asp	Gly	Glu	Trp 180	Ala	His	Glu	Leu	Ala 185	Leu	Asn	Asn	Thr	Val 190	Leu	Glu
Asn	Leu	Asn 195	Phe	Tyr	Met	Thr	Asp 200	Leu	Leu	Gln	Val	Arg 205	Ala	Glu	Asp
Leu	Glu 210	Leu	Ile	Ala	Arg	Asn 215	Cys	Lys	Ser	Leu	Val 220	Ser	Met	Lys	Ile
Ser 225	Glu	Сув	Glu	Ile	Thr 230	Asn	Leu	Leu	Gly	Phe 235	Phe	Arg	Ala	Ala	Ala 240
Ala	Leu	Glu	Glu	Phe 245	Gly	Gly	Gly	Ala	Phe 250	Asn	Asp	Gln	Pro	Glu 255	Leu
Val	Val	Glu	Asn 260	Gly	Tyr	Asn	Glu	His 265	Ser	Gly	ГÀв	Tyr	Ala 270	Ala	Leu
Val	Phe	Pro 275	Pro	Arg	Leu	CÀa	Gln 280	Leu	Gly	Leu	Thr	Tyr 285	Leu	Gly	Arg
Asn	Glu 290	Met	Ser	Ile	Leu	Phe 295	Pro	Ile	Ala	Ser	Arg 300	Leu	Arg	ГÀв	Leu
Asp 305	Leu	Leu	Tyr	Ala	Leu 310	Leu	Asp	Thr	Ala	Ala 315	His	Cys	Phe	Leu	Leu 320
Gln	Arg	Сув	Pro	Asn 325	Leu	Glu	Ile	Leu	Glu 330	Thr	Arg	Asn	Val	Val 335	Gly
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Leu	Arg	Ile 355	Glu	Arg	Gly	Ala	Asp 360	Asp	Gln	Glu	Met	Glu 365	Asp	Glu	Glu
Gly	Ala 370	Val	Thr	His	Arg	Gly 375	Leu	Ile	Asp	Leu	Ala 380	Lys	Gly	CAa	Leu
Glu 385	Leu	Glu	Tyr	Met	Ala 390		Tyr	Val	Ser	Asp 395		Thr	Asn	Glu	Ala 400
Leu	Glu	Val	Ile	Gly 405	Thr	Tyr	Leu	Lys	Asn 410	Leu	Ser	Asp	Phe	Arg 415	Leu
Val	Leu	Leu	Asp 420	Arg	Glu	Glu	Arg	Ile 425	Thr	Asp	Leu	Pro	Leu 430	Asp	Asn
Gly	Val	Arg 435	Ala	Leu	Leu	Arg	Gly 440	Cys	His	Asn	Leu	Arg 445	Arg	Phe	Ala
Leu	Tyr 450	Val	Arg	Pro	Gly	Gly 455	Leu	Thr	Asp	Val	Gly 460	Leu	Ser	Tyr	Val
Gly 465	Gln	Tyr	Ser	Pro	Asn 470	Val	Arg	Trp	Met	Leu 475	Leu	Gly	Tyr	Val	Gly 480
Glu	Ser	Asp	His	Gly 485	Leu	Leu	Glu	Phe	Ser 490	Lys	Gly	Cys	Pro	Ser 495	Leu
Gln	Lys	Leu	Glu 500	Val	Arg	Gly	Сув	Сув 505	Phe	Ser	Glu	Arg	Ala 510	Leu	Ala
Leu	Ala	Thr 515	Leu	Gln	Leu	Lys	Ser 520	Leu	Arg	Tyr	Leu	Trp 525	Val	Gln	Gly

Tyr Arg Ala Ser Ser Ala Gly Arg Asp Leu Leu Ala Met Ala Arg Pro Phe Trp Asn Ile Glu Leu Ile Pro Ala Arg Arg Val Ile Ala Asn Asp Gly Asn Asn Ala Glu Thr Val Val Ser Glu His Pro Ala His Ile Leu Ala Tyr Tyr Ser Leu Ala Gly Gln Arg Thr Asp Phe Pro Asp Thr Val Lys Pro Leu Asp Pro Thr Tyr Leu Leu Ala Glu <210> SEQ ID NO 19 <211> LENGTH: 598 <212> TYPE: PRT <213> ORGANISM: Vitis vinifera <400> SEQUENCE: 19 Met Glu Asp Gly Asn Glu Arg Lys Val Ser Arg Glu Met Leu Asp Met Ala Asp Arg Gly Met Ser Asp Glu Val Leu Asn Cys Val Met Pro Tyr Ile His Asp Pro Lys Asp Arg Asp Ala Val Ser Leu Val Cys Arg Arg 35  $\phantom{\bigg|}40\phantom{\bigg|}40\phantom{\bigg|}45\phantom{\bigg|}$ Trp Tyr Glu Leu Asp Ala Leu Thr Arg Lys His Ile Thr Ile Ala Leu Cys Tyr Thr Thr Thr Pro Gly Arg Leu Arg Gly Arg Phe Pro His Leu 65 70 75 80 Glu Ser Leu Lys Leu Lys Gly Lys Pro Arg Ala Ala Met Phe Asn Leu Ile Met Glu Asp Trp Gly Gly Tyr Val Thr Pro Trp Val Lys Glu Ile 105 Ser Asp Tyr Phe Asp Cys Leu Lys Ser Leu His Phe Arg Arg Met Ile Val Lys Asp Ser Asp Leu Gln Leu Leu Ala Gln Ala Arg Gly Arg Val Leu Leu Val Leu Lys Leu Asp Lys Cys Ser Gly Phe Ser Thr Asp Gly Leu Leu His Val Gly Arg Ser Cys Arg Asn Leu Arg Thr Leu Phe Leu Glu Glu Ser Gln Ile Val Asp Lys Asp Gly Glu Trp Leu His Glu Leu Ala Met Asn Asn Thr Val Leu Glu Thr Leu Asn Phe Tyr Met Thr Glu Leu Ala Thr Val Gln Phe Glu Asp Leu Glu Leu Ile Ala Arg Asn Cys Arg Ser Leu Thr Ser Met Lys Ile Ser Asp Phe Glu Ile Leu Asp Leu 230 Val Gly Phe Phe Arg Ala Ala Thr Ala Leu Glu Glu Phe Ala Gly Gly 250 Ser Phe Ser Glu Gln Ser Asp Lys Tyr Ser Ala Val Ser Phe Pro Pro Lys Leu Cys Arg Leu Gly Leu Asn Tyr Met Gly Lys Asn Glu Met Pro Ile Val Phe Pro Phe Ala Ser Leu Leu Lys Lys Leu Asp Leu Leu Tyr

_	290					295					300				
Crra		T 011	7 an	The	C1.,		IIi a	Ct.a	T 011	Lou		Cl.	T * * * *	Crra	Dwo
305	Leu	ьец	Asp	Inr	310	Asp	HIS	сув	ьeu	315	ше	GIN	гув	сув	320
Asn	Leu	Glu	Phe	Leu 325	Glu	Ala	Arg	Asn	Val 330	Ile	Gly	Asp	Arg	Gly 335	Leu
Glu	Val	Leu	Ala 340	Gln	Ser	CAa	Lys	Lys 345	Leu	Arg	Arg	Leu	Arg 350	Ile	Glu
Arg	Gly	Ala 355	Asp	Glu	Gln	Glu	Met 360	Glu	Asp	Glu	Glu	Gly 365	Val	Val	Ser
Gln	Arg 370	Gly	Leu	Met	Ala	Leu 375	Ala	Arg	Gly	Cys	Leu 380	Glu	Ile	Glu	Tyr
Val 385	Ala	Ile	Tyr	Val	Ser 390	Asp	Ile	Thr	Asn	Ala 395	Ala	Leu	Glu	Càa	Ile 400
Gly	Ala	His	Ser	Lys 405	Lys	Leu	Cys	Asp	Phe 410	Arg	Leu	Val	Leu	Leu 415	Glu
Arg	Glu	Glu	Arg 420	Ile	Thr	Asp	Leu	Pro 425	Leu	Asp	Asn	Gly	Val 430	Arg	Ala
Leu	Leu	Arg 435	Gly	Cys	Gln	Lys	Leu 440	Arg	Arg	Phe	Ala	Leu 445	Tyr	Leu	Arg
Ser	Gly 450	Gly	Leu	Thr	Asp	Val 455	Gly	Leu	Asn	Tyr	Ile 460	Gly	Gln	Tyr	Ser
Pro 465	Asn	Val	Arg	Trp	Met 470	Leu	Leu	Gly	Tyr	Val 475	Gly	Glu	Ser	Asp	Ala 480
Gly	Leu	Leu	Glu	Phe 485	Ser	Arg	Gly	CAa	Pro 490	Ser	Leu	Gln	Lys	Leu 495	Glu
Met	Arg	Gly	500	CAa	Phe	Ser	Glu	Arg 505	Ala	Leu	Ala	Val	Ala 510	Ala	Met
Gln	Leu	Thr 515	Ser	Leu	Arg	Tyr	Leu 520	Trp	Val	Gln	Gly	Tyr 525	Arg	Ala	Ser
Glu	Thr 530	Gly	Arg	Asp	Leu	Leu 535	Val	Met	Ala	Arg	Pro 540	Phe	Trp	Asn	Ile
Glu 545	Leu	Ile	Pro	Ser	Arg 550	Gly	Val	Thr	Ile	Asn 555	Ala	Pro	Asp	Arg	Glu 560
Pro	Val	Ser	Ile	Glu 565	His	Pro	Ala	His	Ile 570	Leu	Ala	Tyr	Tyr	Ser 575	Leu
Ala	Gly	Pro	Arg 580	Thr	Asp	Phe	Pro	Ser 585	Thr	Val	Thr	Pro	Leu 590	Asp	Pro
Ala	Ser	Phe 595	Leu	Thr	Leu										
	D> SI L> LI														
	2 > T? 3 > OI			Pop	ılus	tri	choca	arpa							
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Ile	Ala	Phe		Tyr	Ser	Thr	Ser 40		Asp	Arg	Leu	Arg 45		Arg	Phe
Asn	Asp 50		Glu	Ser	Leu			Lys	Gly	Lys			Ala	Ala	Met
	50					55					60				

Phe 65	Phe	Asn	Leu	Ile	Pro 70	Glu	Asp	Trp	Gly	Gly 75	Phe	Val	Thr	Pro	Trp 80
Val	Asn	Glu	Ile	Ala 85	Glu	Ser	Phe	Asn	GÀa 6	Leu	ГÀв	Ser	Leu	His 95	Phe
Arg	Arg	Met	Ile 100	Val	Lys	Asp	Ser	Asp 105	Leu	Glu	Leu	Leu	Ala 110	Arg	Ser
Arg	Gly	Arg 115	Leu	Leu	Gln	Val	Leu 120	Lys	Leu	Asp	Lys	Cys 125	Ser	Gly	Phe
Ser	Thr 130	Asp	Gly	Leu	Ser	His 135	Ile	Gly	Arg	Ser	Cys 140	Arg	Gln	Leu	Arg
Thr 145	Leu	Phe	Leu	Glu	Glu 150	Ser	Ala	Ile	Val	Glu 155	Arg	Asp	Gly	Asp	Trp 160
Leu	His	Glu	Leu	Ala 165	Thr	Asn	Asn	Thr	Val 170	Leu	Glu	Thr	Leu	Asn 175	Phe
Tyr	Met	Thr	Glu 180	Leu	Thr	Arg	Val	Arg 185	Ser	Glu	Asp	Leu	Glu 190	Leu	Leu
Ala	Arg	Asn 195	CÀa	Arg	Ser	Leu	Val 200	Ser	Val	Lys	Val	Ser 205	Asp	CÀa	Glu
Ile	Leu 210	Asp	Leu	Val	Gly	Phe 215	Phe	His	Ala	Ala	Ser 220	Ala	Leu	Glu	Glu
Phe 225	Cys	Gly	Gly	Ser	Phe 230	Asn	Glu	Pro	Asp	Glu 235	Pro	Asp	Lys	Tyr	Ser 240
Ala	Val	Lys	Phe	Pro 245	Pro	Lys	Leu	Сув	Сув 250	Leu	Gly	Leu	Ser	Tyr 255	Met
Glu	Lys	Asn	Val 260	Met	Ser	Ile	Val	Phe 265	Pro	Phe	Ala	Ser	Leu 270	Leu	Lys
ГÀЗ	Leu	Asp 275	Leu	Leu	Tyr	Ala	Phe 280	Leu	Gly	Thr	Glu	Asp 285	His	Сув	Val
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Ile 305	Gly	Asp	Arg	Gly	Leu 310	Glu	Ala	Leu	Ala	Gln 315	Ser	CAa	Lys	Leu	Leu 320
Lys	Arg	Leu	Arg	Ile 325	Glu	Arg	Gly	Ala	Asp 330	Glu	Gln	Gly	Met	Glu 335	Asp
Val	Asp	Gly	Arg 340	Val	Ser	His	Arg	Gly 345	Leu	Ile	Ala	Leu	Ala 350	Gln	Gly
CÀa	Leu	Glu 355	Leu	Glu	Tyr	Ile	Ala 360	Val	Tyr	Val	Ser	Asp 365	Ile	Thr	Asn
Ala	Ala 370	Leu	Glu	His	Met	Gly 375	Thr	Tyr	Ser	Lys	Asn 380	Leu	Asn	Asp	Phe
Arg 385	Leu	Val	Leu	Leu	Glu 390	Gln	Glu	Glu	Arg	Ile 395	Thr	Asp	Leu	Pro	Leu 400
Asp	Asn	Gly	Val	Arg 405	Ala	Leu	Leu	Arg	Gly 410	Càa	Glu	ГÀв	Leu	Gln 415	Arg
Phe	Gly	Leu	Tyr 420	Leu	Arg	Pro	Gly	Gly 425	Leu	Thr	Asp	Val	Gly 430	Leu	Gly
Tyr	Ile	Gly 435	Gln	Tyr	Ser	Arg	Arg 440	Val	Arg	Trp	Met	Ile 445	Leu	Gly	Ser
Val	Gly 450	Glu	Ser	Asp	Glu	Gly 455	Leu	Leu	Ala	Phe	Ser 460	Arg	Gly	Сув	Pro
Ser 465	Leu	Gln	Lys	Leu	Glu 470	Met	Arg	Ala	Cys	Cys 475	Phe	Ser	Glu	Ser	Ala 480
Leu	Ala	Arg	Ala	Ala	Leu	Gln	Leu	Thr	Ser	Leu	Arg	Tyr	Leu	Trp	Val

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Val	Arg	Pro 515	Phe	Trp	Asn	Ile	Glu 520	Leu	Ile	Pro	Ser	Arg 525	Lys	Val	Glu
Ser	Val 530	Asn	Glu	Ala	Gly	Glu 535	Asn	Ile	Val	Ser	Glu 540	Asn	Pro	Ala	His
Ile 545	Leu	Ala	Tyr	Tyr	Ser 550	Leu	Ala	Gly	Pro	Arg 555	Thr	Asp	Phe	Pro	Asp 560
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Ile	Ala	Leu 35	CÀa	Tyr	Ser	Thr	Ser 40	Pro	Asp	Arg	Leu	Gln 45	Arg	Arg	Phe
ГÀа	His 50	Leu	Glu	Ser	Leu	Lys 55	Met	Lys	Gly	Lys	Pro 60	Arg	Ala	Ala	Met
Phe 65	Phe	Asn	Leu	Ile	Pro 70	Asp	Asp	Trp	Gly	Gly 75	Phe	Val	Thr	Pro	Trp 80
Val	Asn	Glu	Ile	Ala 85	Glu	Ser	Phe	Asn	oo Oys	Leu	ГÀз	Ser	Leu	His 95	Phe
Arg	Arg	Met	Ile 100	Val	ràa	Asp	Ser	Asp 105	Leu	Glu	Leu	Leu	Ala 110	Ser	Ser
Arg	Gly	Lys 115	Val	Leu	Gln	Val	Leu 120	ГÀЗ	Leu	Asp	ГÀЗ	Суs 125	Ser	Gly	Phe
Ser	Thr 130	Asp	Gly	Leu	Ser	His 135	Ile	Gly	Arg	Ser	Cys 140	Arg	Gln	Leu	Arg
Thr 145	Leu	Phe	Leu	Glu	Glu 150	Ser	Ala	Ile	Ala	Tyr 155	Glu	ГÀа	Asp	Gly	Asp 160
Trp	Leu	His	Glu	Leu 165	Ala	Thr	Asn	Asn	Thr 170	Val	Leu	Glu	Thr	Leu 175	Asn
Phe	Tyr	Met	Thr 180	Asp	Leu	Thr	Lys	Val 185	Arg	Leu	Glu	Asp	Leu 190	Glu	Leu
Leu	Ala	Lys 195	Asn	Cys	Arg	Ser	Leu 200	Val	Ser	Val	Lys	Ile 205	Ser	Asp	Cys
Glu	Ile 210	Leu	Glu	Leu	Val	Gly 215	Phe	Phe	Arg	Ala	Ala 220	Ser	Ala	Ile	Glu
Glu 225	Phe	Cys	Gly	Gly	Ser 230	Phe	Asn	Glu	Pro	Asp 235	Gln	Pro	Gly	Lys	Tyr 240
Ser	Ala	Val	Val	Phe 245	Pro	Pro	Lys	Leu	Сув 250	Arg	Leu	Gly	Leu	Ser 255	Tyr
Met	Glu	Lys	Asn 260	Val	Met	Ser	Ile	Val 265	Phe	Pro	Phe	Ala	Ser 270	Leu	Leu
Lys	Lys	Leu 275	Asp	Leu	Leu	Tyr	Val 280	Leu	Leu	Gly	Thr	Glu 285	Asp	His	Cys

Val	Leu 290	Val	Gln	Arg	Cys	Pro 295	Asn	Leu	Glu	Val	Leu 300	Glu	Thr	Arg	Asn
Val 305	Ile	Gly	Asp	Arg	Gly 310	Leu	Glu	Ala	Leu	Ala 315	Arg	Ser	Cys	Lys	Arg 320
Leu	Lys	Arg	Leu	Arg 325	Ile	Glu	Arg	Gly	Ala 330	Asp	Glu	Gln	Glu	Met 335	Glu
Asp	Val	Asp	Gly 340	Arg	Val	Ser	Gln	Arg 345	Gly	Leu	Ile	Ala	Leu 350	Ala	Gln
Gly	Cys	Leu 355	Glu	Leu	Glu	Tyr	Ile 360	Ala	Val	Tyr	Val	Ser 365	Asp	Ile	Ser
Asn	Ala 370	Ala	Leu	Glu	His	Met 375	Gly	Ala	Tyr	Ser	380	Asn	Leu	Asn	Asp
Phe 385	Arg	Leu	Val	Leu	Leu 390	Glu	Gln	Glu	Asp	Arg 395	Ile	Thr	Asp	Leu	Pro 400
Leu	Asp	Asn	Gly	Val 405	Arg	Ala	Leu	Leu	Arg 410	Gly	CÀa	Glu	ГЛа	Leu 415	Gln
Arg	Phe	Gly	Leu 420	Tyr	Leu	Arg	Ser	Gly 425	Gly	Leu	Thr	Asp	Val 430	Gly	Leu
Gly	Tyr	Ile 435	Gly	Gln	Tyr	Ser	Arg 440	His	Val	Arg	Trp	Met 445	Ile	Leu	Gly
Ser	Val 450	Gly	Glu	Ser	Asp	Glu 455	Gly	Leu	Leu	Ala	Phe 460	Ser	Met	Gly	Cys
Pro 465	Ser	Leu	Gln	ГÀа	Leu 470	Glu	Met	Arg	Ala	Cys 475	CÀa	Phe	Thr	Glu	Arg 480
Ala	Leu	Ala	Arg	Ala 485	Ala	Leu	Gln	Leu	Thr 490	Ser	Leu	Arg	Tyr	Leu 495	Trp
Val	His	Gly	Tyr 500	Arg	Glu	Thr	Ser	Asn 505	Gly	His	Arg	Asp	Leu 510	Leu	Thr
Met	Val	Arg 515	Pro	Phe	Trp	Asn	Ile 520	Glu	Leu	Ile	Pro	Ser 525	Arg	Arg	Val
Ala	Thr 530	Val	Asn	Asn	Ala	Gly 535	Glu	Asp	Ile	Val	Ser 540	Glu	Asn	Pro	Ala
His 545	Ile	Leu	Ala	Tyr	Tyr 550	Ser	Leu	Ala	Gly	Pro 555	Arg	Thr	Asp	Phe	Pro 560
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Tyr	Ile	Gln 35	Gly	Pro	Lys	Asp	Arg 40	Asp	Ala	Val	Ser	Leu 45	Val	Cys	Arg
Arg	Trp 50	Tyr	Glu	Leu	Asp	Ala 55	Leu	Thr	Arg	Lys	His 60	Ile	Thr	Ile	Ala
Leu 65	Сув	Tyr	Thr	Thr	Ser 70	Pro	Asp	Arg	Leu	Arg 75	Arg	Arg	Phe	Lys	His 80
Leu	Glu	Ser	Leu	Lys 85	Leu	Lys	Gly	Lys	Pro 90	Arg	Ala	Ala	Met	Phe 95	Asn

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Leu	Ile	Pro	Glu 100	Asp	Trp	Gly	Gly	Tyr 105	Val	Thr	Pro	Trp	Ile 110	Asp	Glu
Ile	Ala	Ala 115	Ala	Ser	Phe	Thr	Cys 120	Leu	Lys	Ser	Leu	His 125	Phe	Lys	Arg
Met	Ile 130	Val	Lys	Asp	Ser	Asp 135	Leu	Ala	Leu	Leu	Ala 140	Lys	Ser	Arg	Gly
Lys 145	Val	Leu	His	Val	Leu 150	Lys	Leu	Asp	Lys	Сув 155	Ser	Gly	Phe	Ser	Thr 160
Asp	Gly	Leu	Leu	His 165	Val	Ala	CÀa	Phe	Cys 170	Arg	Gln	Leu	Arg	Thr 175	Leu
Phe	Leu	Glu	Glu 180	Ser	Ala	Ile	Phe	Glu 185	ГЛа	Asp	Gly	Asp	Trp 190	Leu	His
Glu	Ile	Ala 195	Met	Asn	Asn	Thr	Val 200	Leu	Glu	Ile	Leu	Asn 205	Phe	Tyr	Met
Thr	Asp 210	Leu	Asn	Ala	Val	Arg 215	Phe	Glu	Asp	Leu	Glu 220	Ile	Ile	Ala	Lys
Asn 225	Cys	Arg	Cys	Leu	Val 230	Ser	Val	Lys	Ile	Ser 235	Asp	CAa	Glu	Ile	Leu 240
Asp	Leu	Ala	Gly	Phe 245	Phe	His	Ala	Ala	Ala 250	Ala	Leu	Glu	Glu	Phe 255	Cya
Gly	Gly	Ser	Phe 260	Asn	Tyr	Ser	Ala	Asn 265	Asp	Leu	Gln	Asp	Lys 270	Tyr	Ser
Ala	Val	Thr 275	Phe	Pro	Arg	ГЛа	Leu 280	Cha	Arg	Leu	Gly	Leu 285	Thr	Tyr	Leu
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Lys 305	Leu	Asp	Leu	Leu	Tyr 310	Ala	Leu	Leu	Asp	Thr 315	Glu	Asp	His	CAa	Leu 320
Leu	Ile	Gln	ГÀз	Phe 325	CAa	Asn	Leu	Glu	Val 330	Leu	Glu	Thr	Arg	Asn 335	Val
Ile	Gly	Asp	Arg 340	Gly	Leu	Glu	Val	Leu 345	Ala	Ser	Ser	CAa	Lys 350	Arg	Leu
Lys	Arg	Leu 355	Arg	Ile	Glu	Arg	Gly 360	Ala	Asp	Glu	Gln	Gly 365	Met	Glu	Asp
Glu	Glu 370	Gly	Ile	Val	Ser	His 375	Arg	Gly	Leu	Ile	Ala 380	Leu	Ala	Gln	Gly
Сув 385	Leu	Glu	Leu	Glu	Tyr 390	Leu	Ala	Val	Tyr	Val 395	Ser	Asp	Ile	Thr	Asn 400
Ala	Ala	Leu	Glu	His 405	Ile	Gly	Ala	His	Leu 410	Lys	Asn	Leu	Asn	Asp 415	Phe
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Phe	Ala 450	Leu	Tyr	Leu	Arg	Pro 455	Gly	Gly	Leu	Thr	Asp 460	Val	Gly	Leu	Gly
Tyr 465	Ile	Gly	Glu	Tyr	Ser 470	Pro	Asn	Val	Arg	Trp 475	Met	Leu	Leu	Gly	Tyr 480
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### -continued

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Ile Ala	. Lys	Сув	Pro 325	Asn	Leu	Leu	Val	Leu 330	Ala	Val	Arg	Asn	Val 335	Ile
Gly As	Arg	Gly 340	Leu	Gly	Val	Val	Ala 345	Asp	Thr	Cys	Lys	Lys 350	Leu	Gln
Arg Let	ı Arg 355	Ile	Glu	Arg	Gly	Asp 360	Asp	Glu	Gly	Gly	Val 365	Gln	Glu	Glu
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Arg Glu 385	ı Leu	Glu	Tyr	Ile 390	Ala	Ala	Tyr	Val	Ser 395	Asp	Ile	Thr	Asn	Gly 400
Ala Le	ı Glu	Ser	Ile 405	Gly	Thr	Phe	Cys	Lys 410	Lys	Leu	Tyr	Asp	Phe 415	Arg
Leu Val	Leu	Leu 420	Asp	Arg	Glu	Glu	Arg 425	Ile	Thr	Asp	Leu	Pro 430	Leu	Asp
Asn Gly	7 Val 435	Arg	Ala	Leu	Leu	Arg 440	Gly	Cys	Thr	Lys	Leu 445	Arg	Arg	Phe
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Gly Gl	1 Thr	Asp	Asp 485	Gly	Leu	Ile	Ser	Phe 490	Ala	Leu	Gly	CÀa	Val 495	Asn
Leu Arç	d Tàs	Leu 500	Glu	Leu	Arg	Ser	Cys	Сув	Phe	Ser	Glu	Arg 510	Ala	Leu
Ala Le	1 Ala 515	Ile	Leu	His	Met	Pro 520	Ser	Leu	Arg	Tyr	Val 525	Trp	Val	Gln
Gly Tyr		Ala	Ser	Gln	Thr 535	Gly	Arg	Asp	Leu	Met 540	Leu	Met	Ala	Arg
Pro Phe	e Trp	Asn	Ile	Glu 550	Phe	Thr	Pro	Pro	Asn 555	Pro	Lys	Asn	Gly	Gly 560
Trp Let	ı Met	Glu	Asp 565	Gly	Glu	Pro	Cys	Val 570	Asp	Ser	His	Ala	Gln 575	Ile
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Ser Gly	/ Met	Ser 20	Asp	Val	Val	Leu	Gly 25	Сув	Val	Met	Pro	Tyr 30	Ile	His
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Glu Le	ı Asp	Ala	Leu	Thr	Arg	Lys	His	Ile	Thr	Ile	Ala	Phe	Cys	Tyr

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Leu 305	Asp	Thr	Glu	Asp	His 310	CÀa	Leu	Leu	Ile	Gln 315	Lys	CAa	Phe	Asn	Leu 320
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Val 385	Tyr	Val	Ser	Asp	Ile 390	Thr	Asn	Ala	Ala	Leu 395	Glu	His	Ile	Gly	Thr 400
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Met	Gln	Arg 435	Lys	Leu	Arg	Arg	Phe 440	Ala	Leu	Tyr	Leu	Arg 445	Pro	Gly	Gly
Leu	Thr 450	Asp	Glu	Gly	Leu	Gly 455	Tyr	Ile	Gly	Gln	His 460	Ser	Lys	Asn	Val
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Thr 305	Glu	Asp	His	CÀa	Thr 310	Leu	Ile	Gly	Lys	Cys 315	Pro	Asn	Leu	Glu	Ile 320
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475 Tyr Val Gly Glu Ser Asp Glu Gly Leu Leu Lys Phe Leu Lys Asp Asp Glu Gly Form Ser		450					455					460				
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Leu Ala Leu Ala Ala Met Gln Leu Lys Ser Leu Arg Tyr Leu Trp 515 515 515 515 515 515 515 515 515 51	Tyr	Val	Gly	Glu		Asp	Glu	Gly	Leu		Lys	Phe	Leu	Lys	_	Val
S15	Leu	Thr	Сув		Ala	Arg	Ser	Glu		Leu	Leu	Phe	Ser		Arg	Ala
530 535 540 540 540 545 540 545 546 545 546 545 545 546 545 545 545	Leu	Ala		Ala	Ala	Met	Gln		Lys	Ser	Leu	Arg		Leu	Trp	Val
550 555  Ser Glu Gly Asn Asn Gly Glu Thr Ile Val Ala Glu His Pro Ala 575  Ile Leu Ala Tyr Tyr Ser Leu Ala Gly Gln Arg Thr Asp Phe Pro 585  Thr Val Arg Pro Leu Asp Pro Asn Ser Leu Leu Ala Glu 605  **C110> SEQ ID NO 27	Gln		Tyr	Arg	Ala	Ser		Ala	Gly	Arg	Asp		Leu	Ala	Met	Ala
Septended   Sept		Pro	Phe	Trp	Asn		Glu	Leu	Ile	Pro		Arg	Arg	Val	Val	Ser 560
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	Gly			245			-	_	250					255	_
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Gly	Leu																

What is claimed is:

- 1. A method for targeted protein degradation in a cultured yeast or mammalian host cell comprising:
  - a) introducing a DNA sequence encoding a target protein tagged with one or more peptide tags into said host cell, wherein said peptide tags consist of the amino acid sequence as set forth in SEQ ID NOs: 5, 6 or 7;
  - b) introducing a DNA sequence encoding *Arabidopsis* protein COI1 or a homolog thereof into said host cell, wherein said *Arabidopsis* protein COI1 or a homolog thereof comprises the amino acid sequence as set forth in SEQ ID NOs: 15-27 or 28;
  - c) culturing said host cell under conditions that result in expression of said tagged target protein and said *Arabidopsis* protein COII or a homolog thereof; and
  - d) contacting said host cell from step (c) with a molecule that binds the COI1/jasmonyl-L-isoleucine (JA-Ile) binding pocket of COI1, wherein said molecule is selected from the group consisting of coronatine and JA-Ile, and wherein contacting of said molecule results in degradation of said tagged target protein.
- **2**. A method for targeted protein degradation in a cultured yeast or mammalian host cell comprising:

- a) introducing a DNA sequence encoding one or more peptide tags into said host cell adjacent to a DNA sequence encoding an endogenous target protein, wherein said peptide tags consist of the amino acid sequence as set forth in SEQ ID NOs:5, 6 or 7;
- b) introducing a DNA sequence encoding *Arabidopsis* protein COI1 or a homolog thereof into said host cell, wherein said *Arabidopsis* protein COI1 or a homolog thereof comprises the amino acid sequence as set forth in SEQ ID NOs: 15-27 or 28;
- c) culturing said host cell under conditions that result in expression of said endogenous target protein tagged with said one or more peptide tags and said *Arabidopsis* protein COI1 or a homolog thereof; and
- d) contacting said host cell of step (c) with a molecule that binds the COI1/jasmonyl-L-isoleucine (JA-Ile) binding pocket of COI1, wherein said molecule is selected from the group consisting of coronatine and JA-Ile, and wherein contacting of said molecule results in degradation of said target protein.
- 3. The method of claim 1 or 2, further comprising the step of contacting said host cell with an inositol pentakisphophate cofactor.

\* \* \* \* \*